

Model Biological Membrane Platform: Current State and Progress

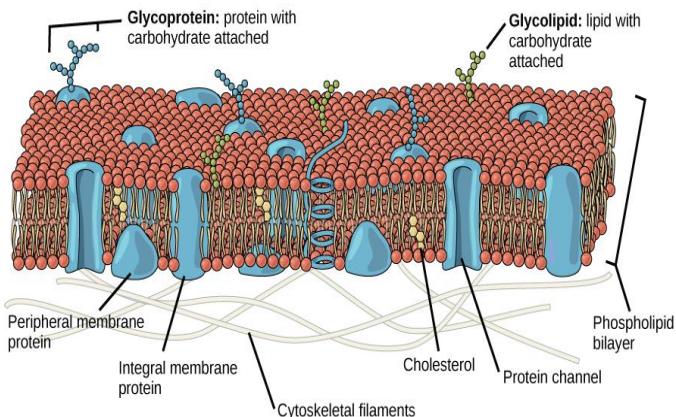
Irena Kiesel

Metting Sacley
28th-29th May 2015

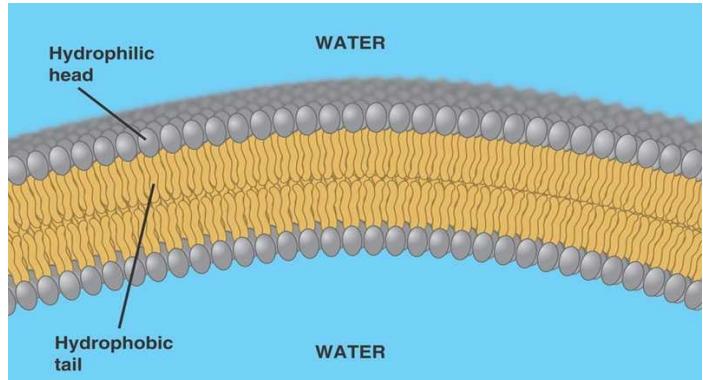


- Deuterated Lipids Extraction (Robin Delhom)
- Polymer Tethered Membranes (Irena Kiesel)
 - Idea
 - Test with commercially available lipids
 - Current state with natural lipids
- Liquid/Liquid interface (Ernesto Scoppola)

Motivation



Real system: complex

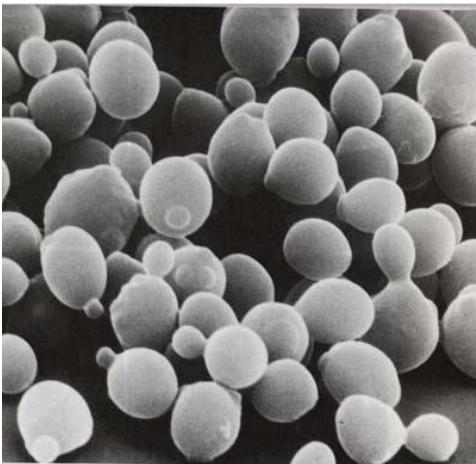


Model system: Mimic real system
with controlled conditions

- Investigate transport processes through membranes (drugs, proteins, diseases)
 - Natural, deuterated lipids
 - Tethered membranes for neutron reflectometry
- Create a recipe for reproducible and stable model membranes

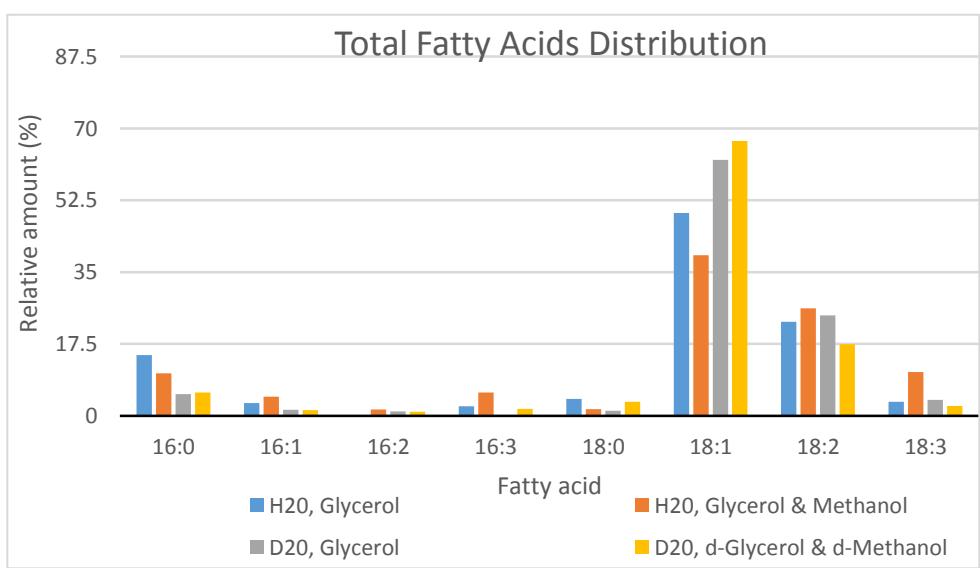
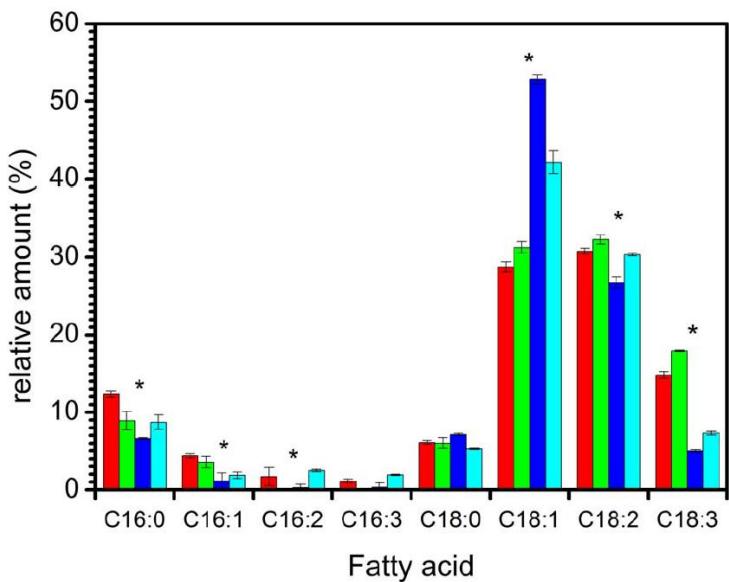
Pichia pastoris yeast

Composition
 h-lipids = d-lipids ?!



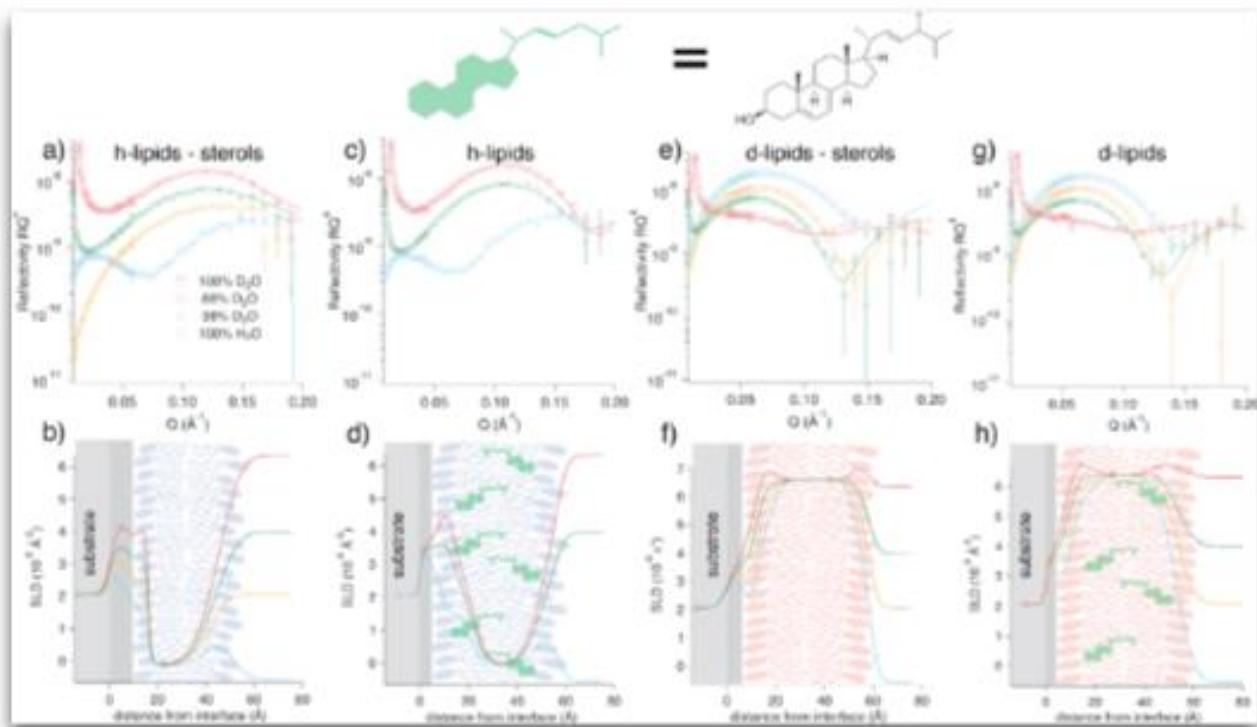
Robin Delhom

Δ Growth temperature Δ Carbon source



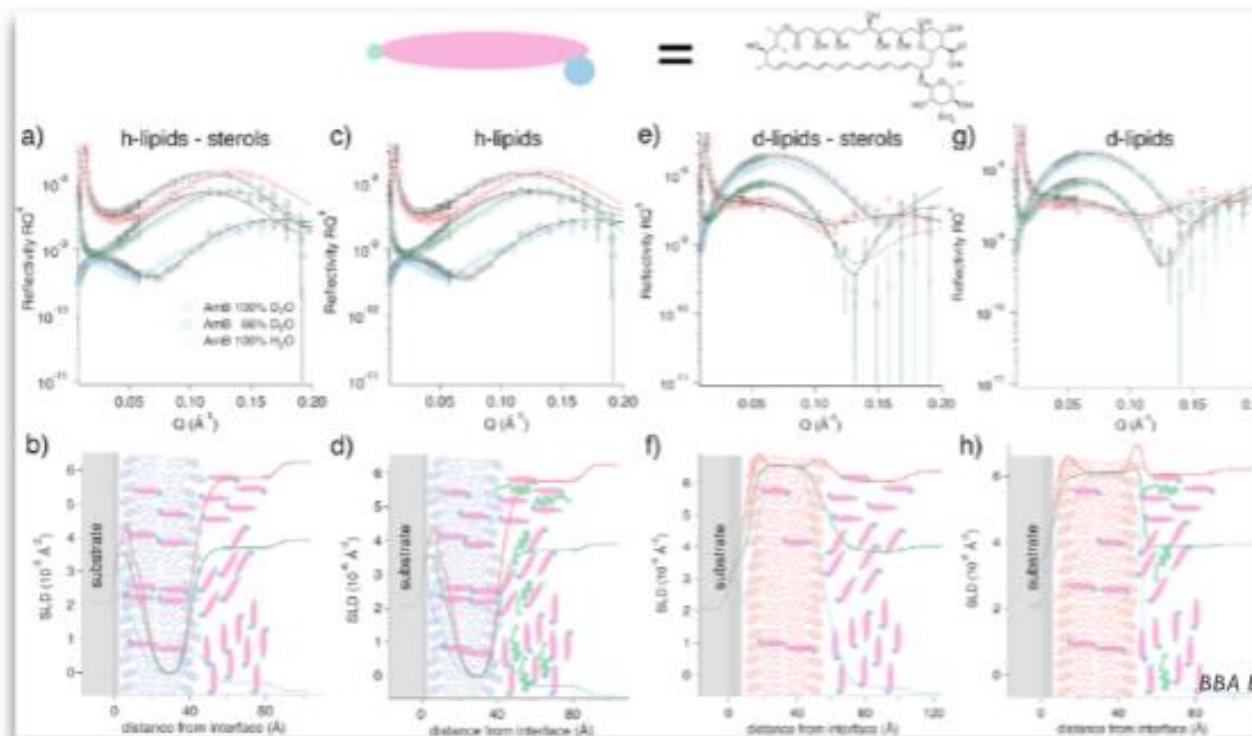
DEUTERATED NATURAL LIPID EXTRACTION - STRUCTURAL CHARACTERISATION AND INTERACTION WITH AN ANTIBIOTIC (SINE2020)

PhD A. de Ghellinck, R. Delhom



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PlosONE 2014

BBA Biomembranes under revision

Characterization by Neutron Diffraction



Q-range $0.02 - 2.5 \text{ \AA}^{-1}$

Times ~min



research papers

Acta Crystallographica Section D
Biological
Crystallography
ISSN 1399-0047

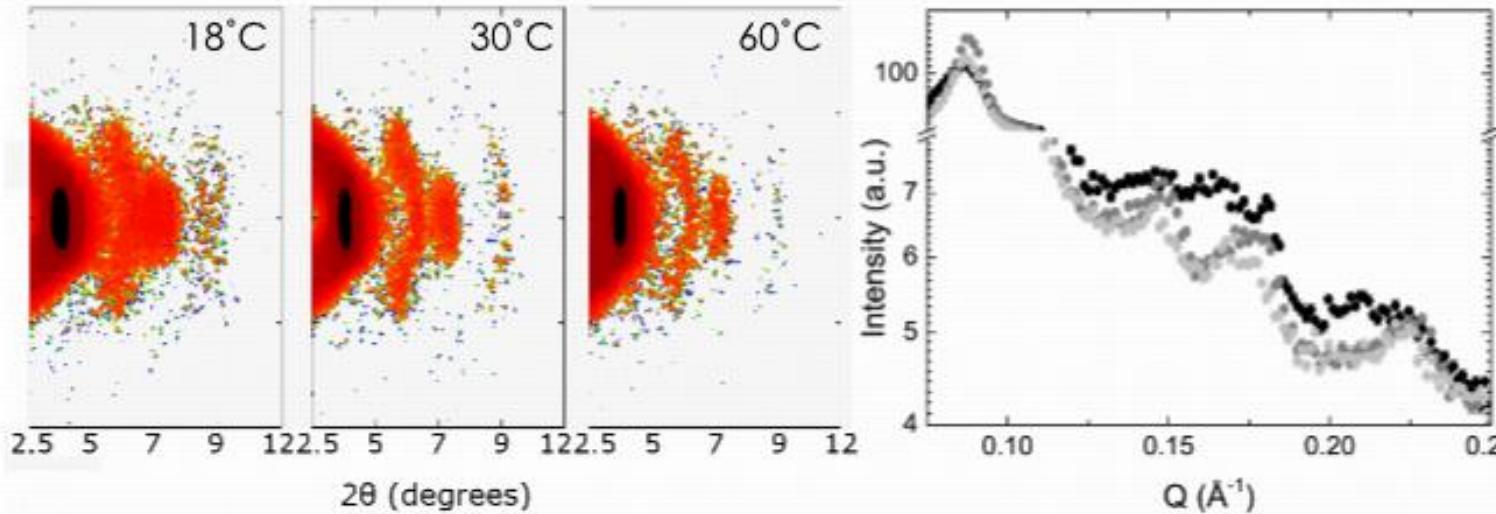
Yuri Gerelli,^{a,*} Alexis de Ghellinck,^{a,b} Juliette Jouhet,^c Valérie Laux,^a Michael Haerlein^a and Giovanna Fragneto^{a*}

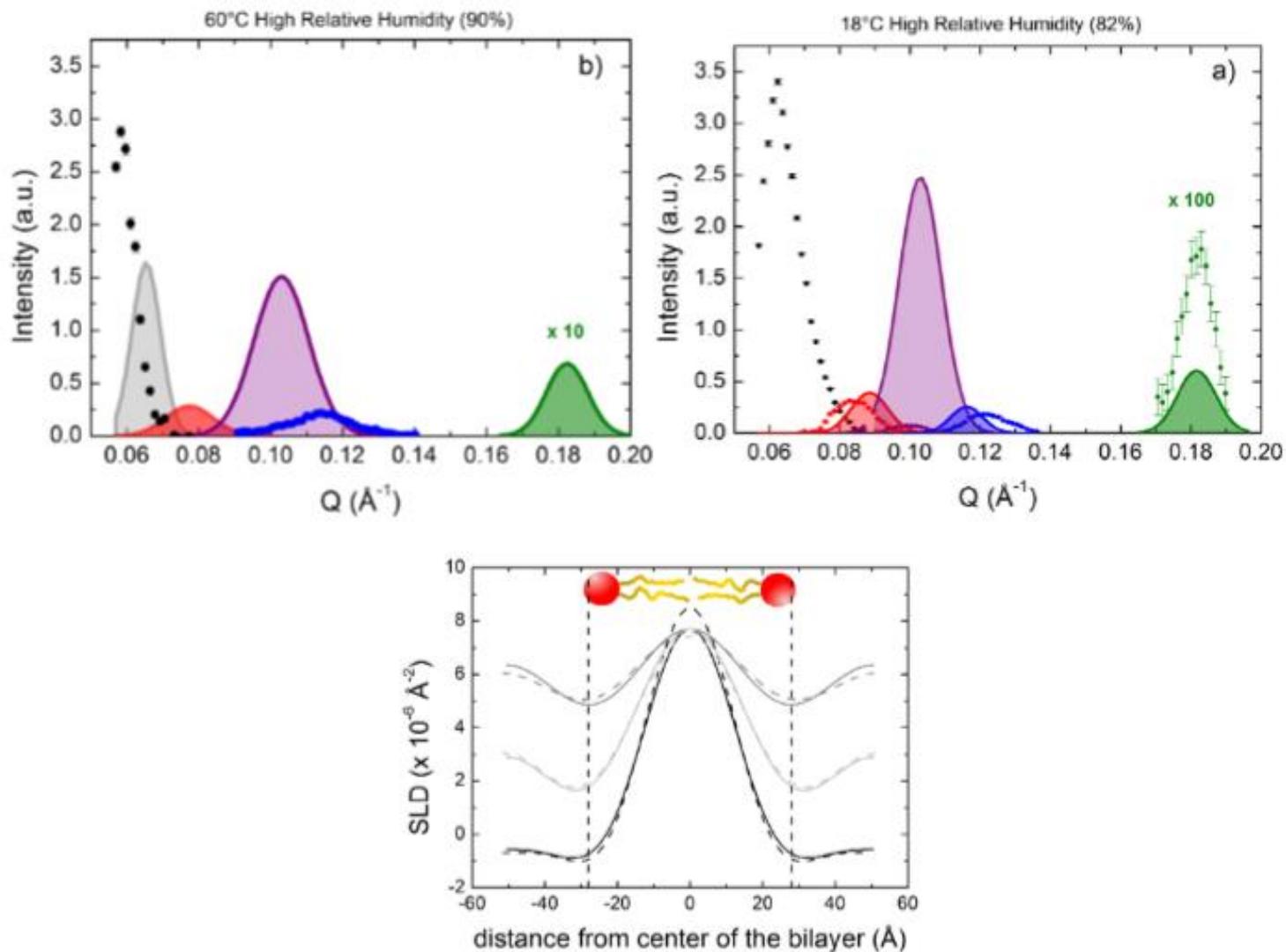
Multi-lamellar organization of fully deuterated lipid extracts of yeast membranes

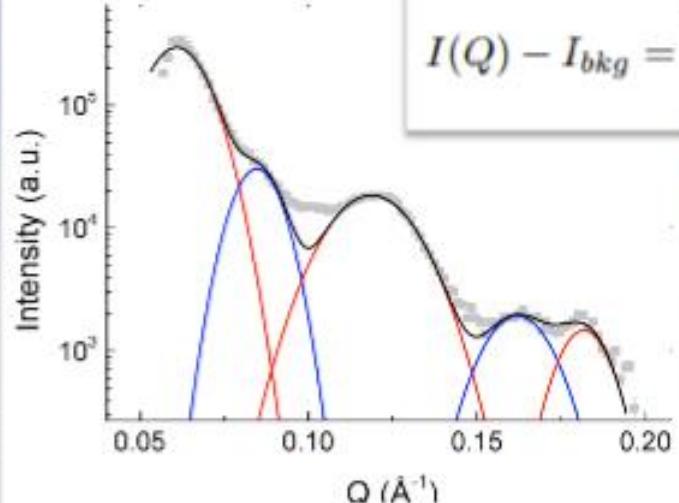
Received 29 April 2014

Accepted 18 October 2014

Neutron scattering studies on mimetic biomembranes are currently limited by the low availability of deuterated unsaturated lipid species. In the present work, results from the first neutron diffraction experiments on fully deuterated lipid extracts from the yeast *Pichia pastoris* are presented. The structural features of these fully deuterated lipid stacks are compared with those of their hydrogenous analogues

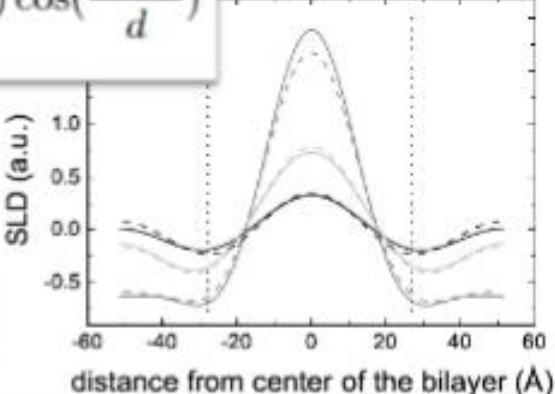






$$I(Q) - I_{bkg} = \sum_{h=1}^{h_{max}} a_h e^{-\frac{1}{2} \left(\frac{Q-Q_h}{\sigma_k} \right)^2}$$

$$\rho(z) = \frac{2}{d} \sum_{h=1}^N F(h) \cos\left(\frac{2\pi h z}{d}\right)$$



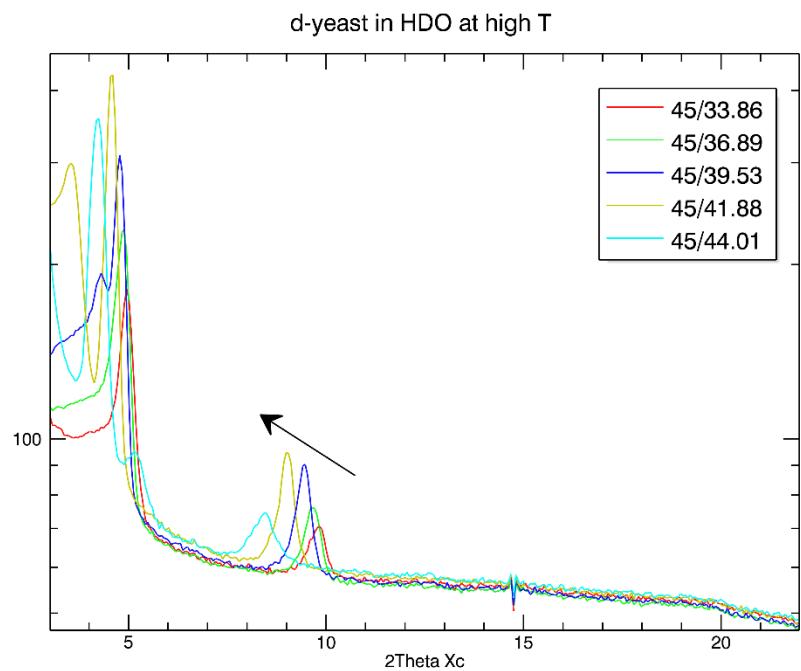
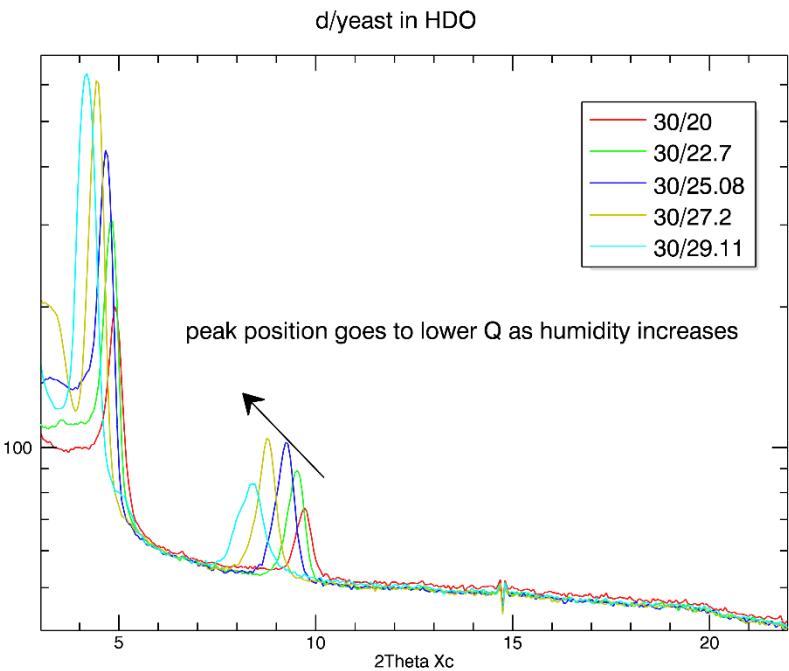
PC	46.8%		16:0	9%
PE	12.9%		18:0	5%
PG ⁻	1.7%		16:1	1%
PI ⁻	12%		18:1	61%
PS ⁻	9.8%		18:2	7%
CL ⁻	4.2%			
Head % chain composition				

	HRH		LRH	
	$d_a (\text{\AA})$	$d_b (\text{\AA})$	$d_a (\text{\AA})$	$d_b (\text{\AA})$
18°C	105 ± 2	76 ± 3	90 ± 10	72 ± 5
30°C	105 ± 2	76 ± 3	86 ± 5	73 ± 2
60°C	104 ± 2	---	82 ± 6	74 ± 2

Few peaks = disordered system

Two d-spacings due to inhomogeneity of sample

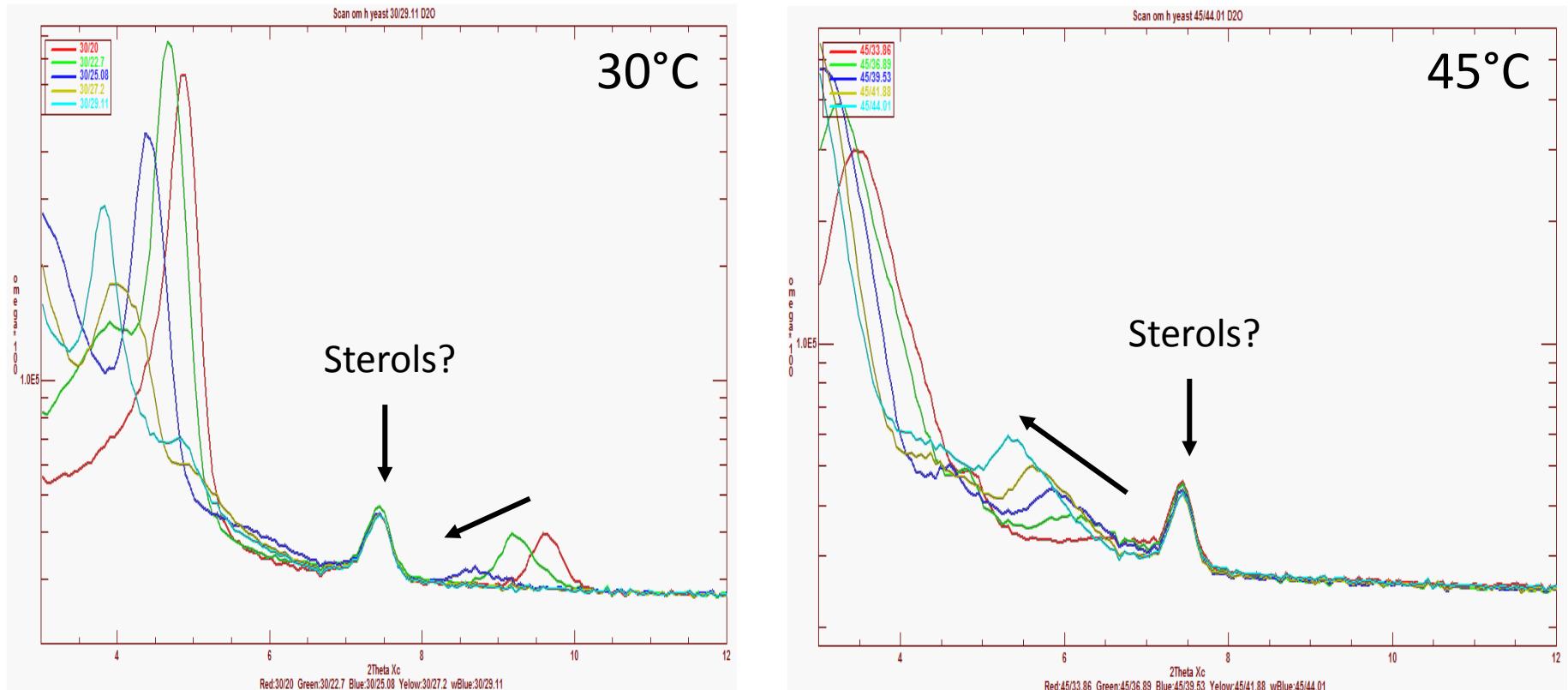
Large d-spacing due to large presence of negatively charged lipids



Spincoated multilayer of deuterated yeast lipids extracts on silicon

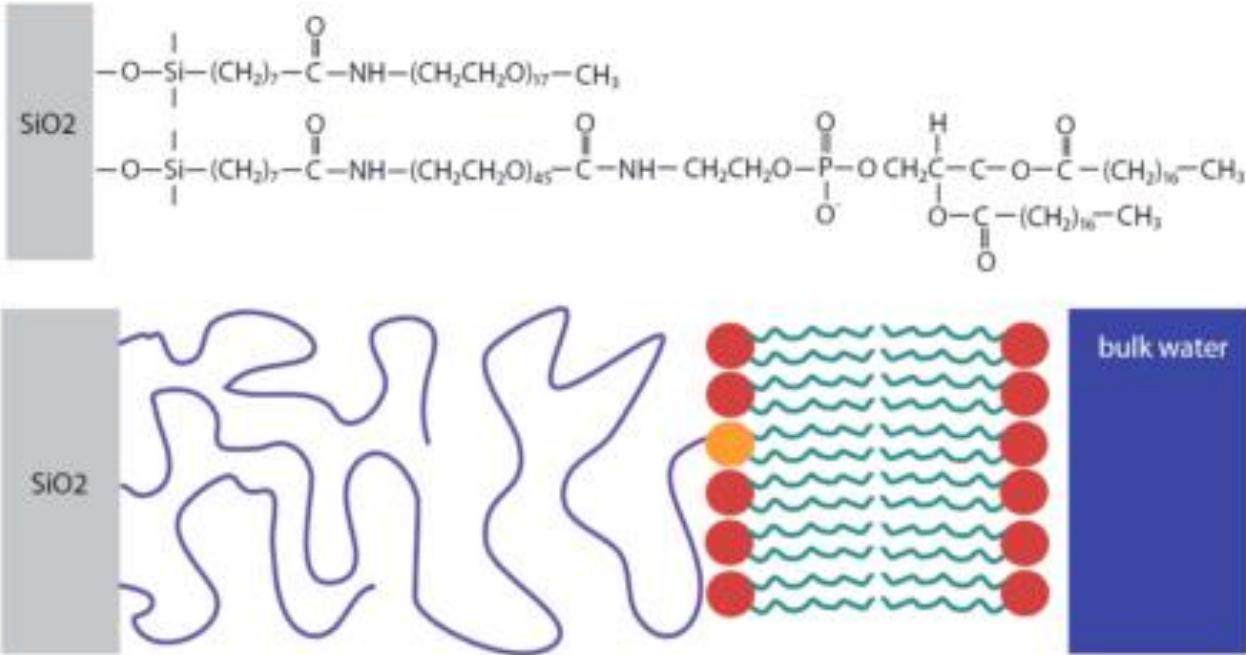
- Humidity chamber
- Different temperatures (30°C and 45°C)
- Gracing incident diffraction:
 - peak → Order of the lipid-stacks, distance in between

Hydrogenated Lipid Extract



Multilayer hYeast at 30°C and 45°C and different relative humidities
Differences between hYeast and dYeast
Analysis in progress

Collaboration with AG Nickel from Munich:
Polymertethered spincoated membranes

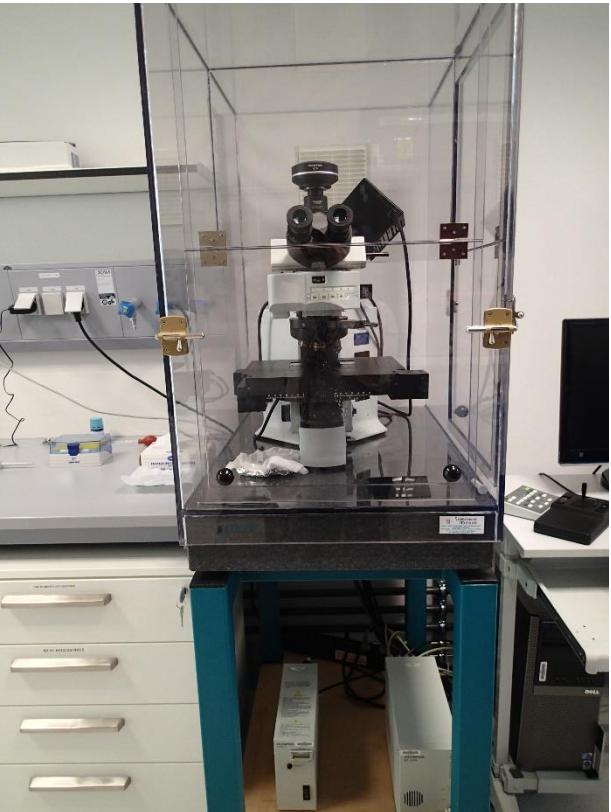


Hertich et al.,
Langmuir 2014, 30, 9442-944

Deuterated and natural lipids extracted from yeast (de Ghellnick,
current project PhD Robin Delhom)

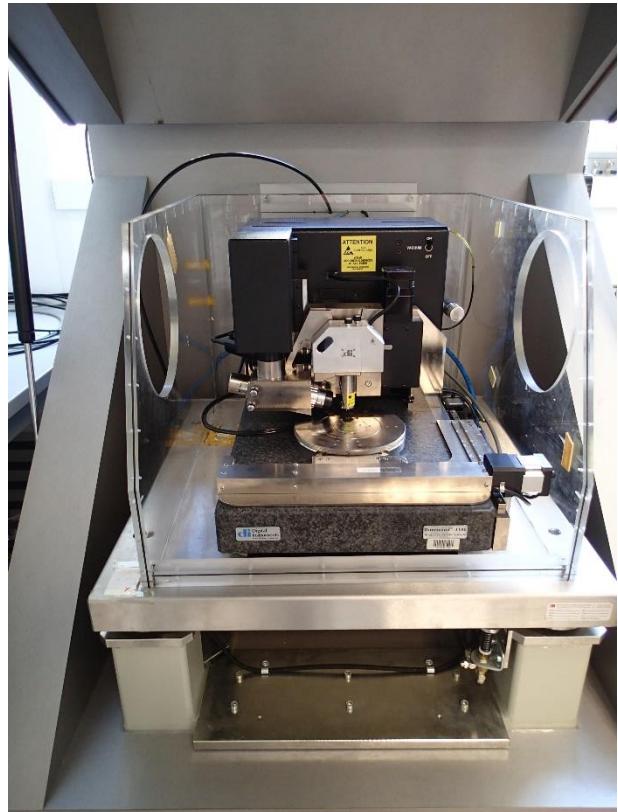
Advantage: - Fast easy sample preparation with spin-coating
 - Reusable substrates

Precharacterisation



Fluorescence Microscopy

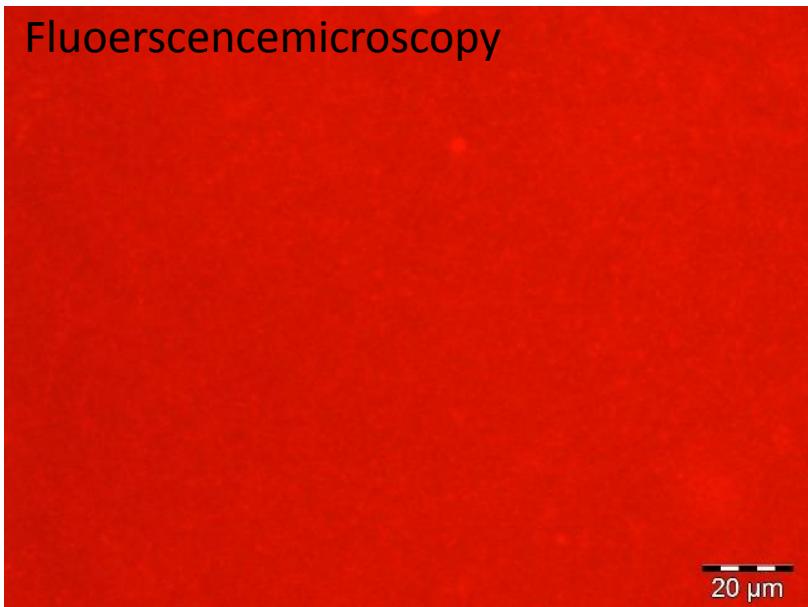
+ Ellipsometer



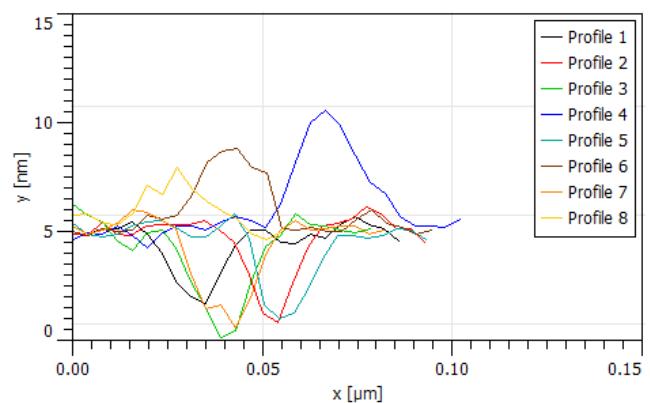
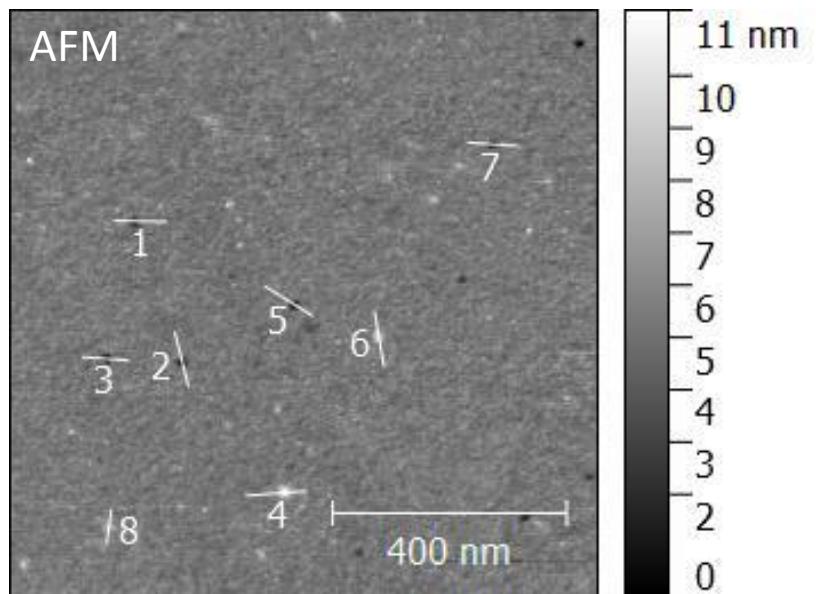
AFM (Veeco)

- Ordered a liquid head for measurements in water

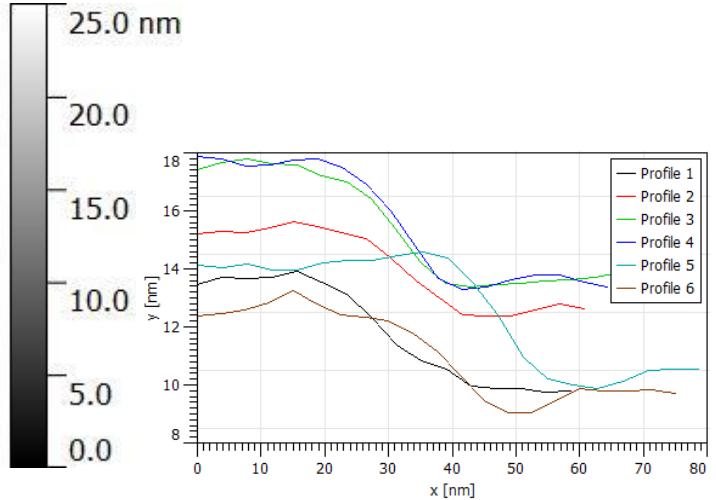
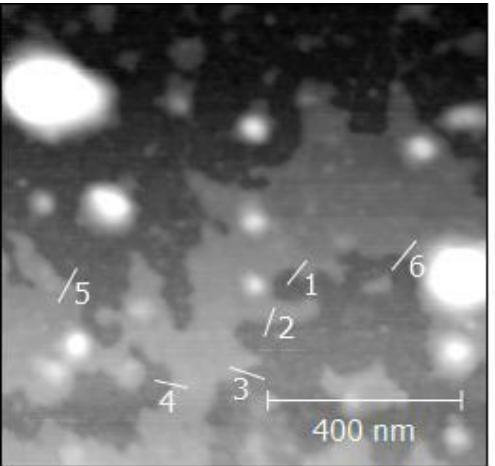
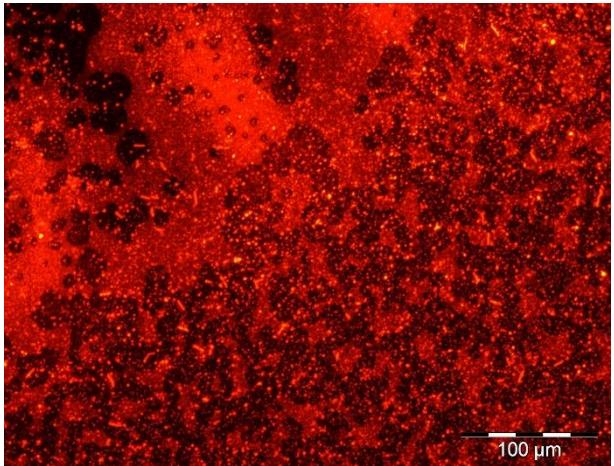
- First substrates successfully prepared
- Test with synthesized lipids (SOPC on PEG/Silicon)
 - Different concentrations



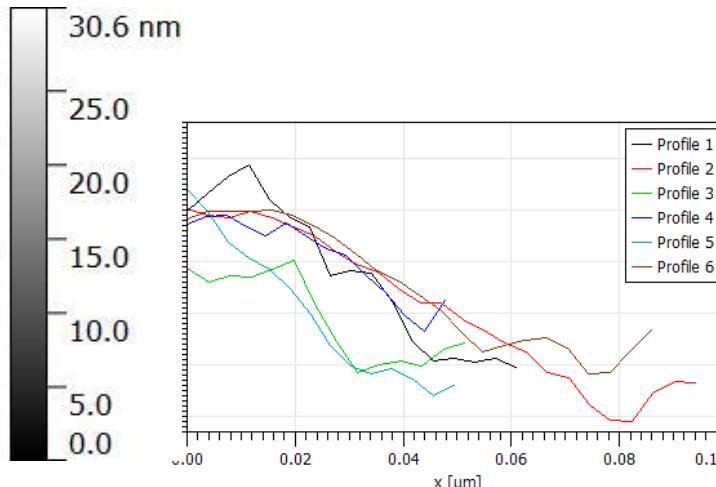
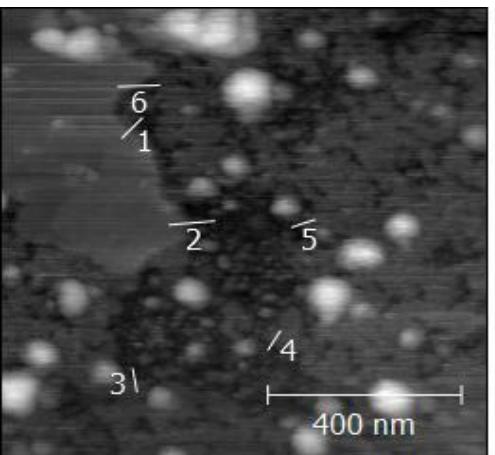
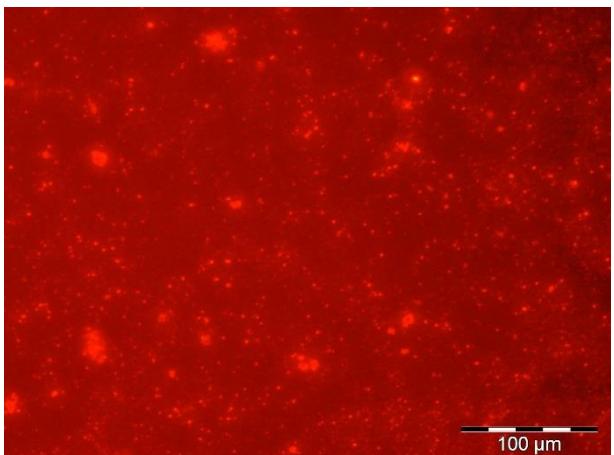
2 mg/mL SOPC/Isopropanol
measurements in water



Natural lipids



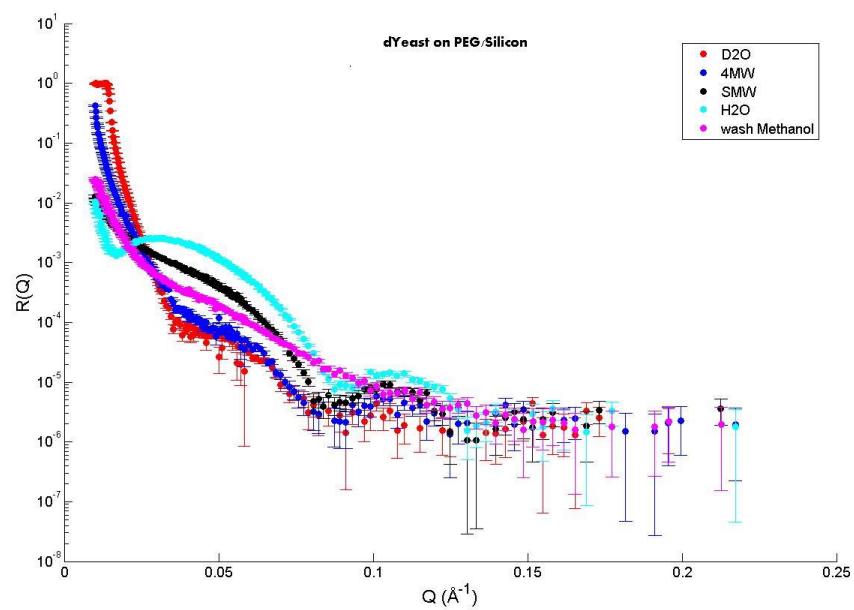
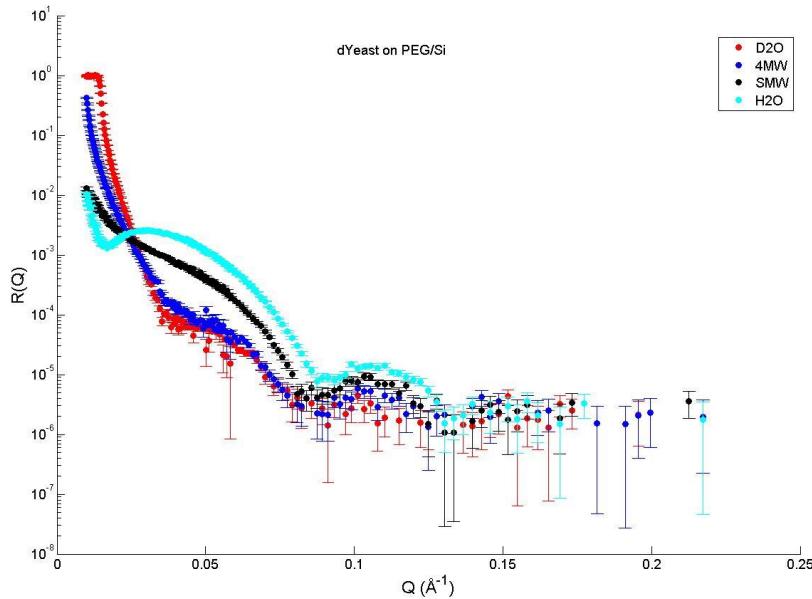
Air



Water

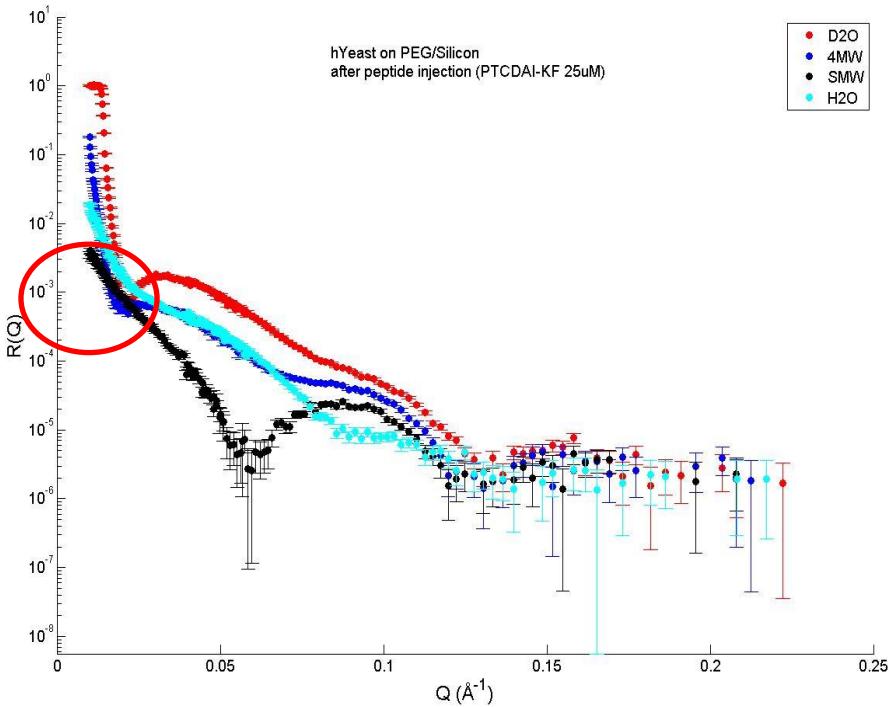
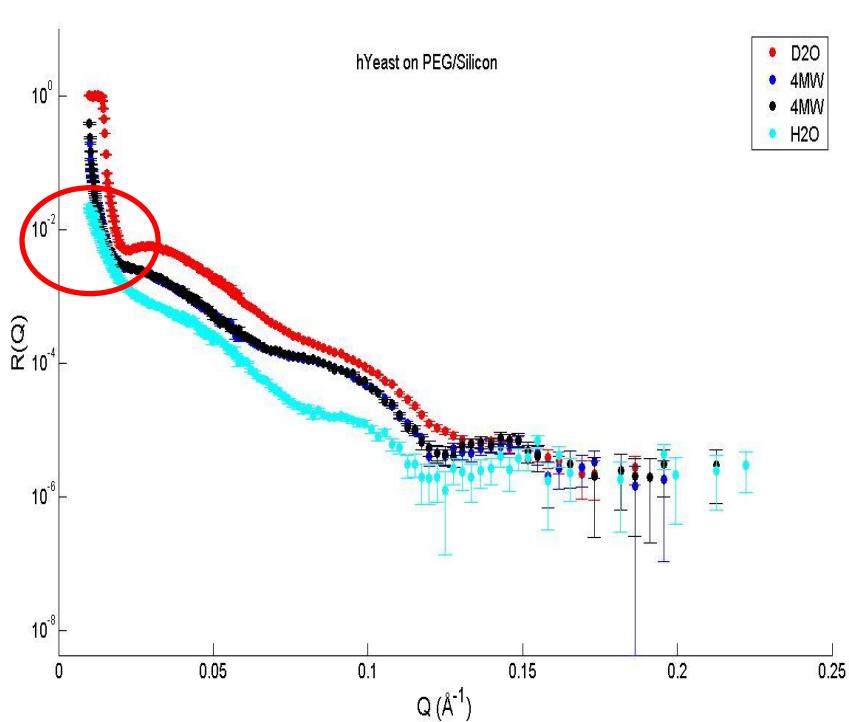
1.5 mg/mL hYeast/Methanol+Chloroform → Solubility

- Natural Yeast lipids, deuterated, spincoated on PEG/Silicon



- Nice features in the reflectivities for different contrasts
- Features fits to the size of the PEG Layer
- Proof, that we can wash away the lipids with methanol
- Analysis is work in progress...

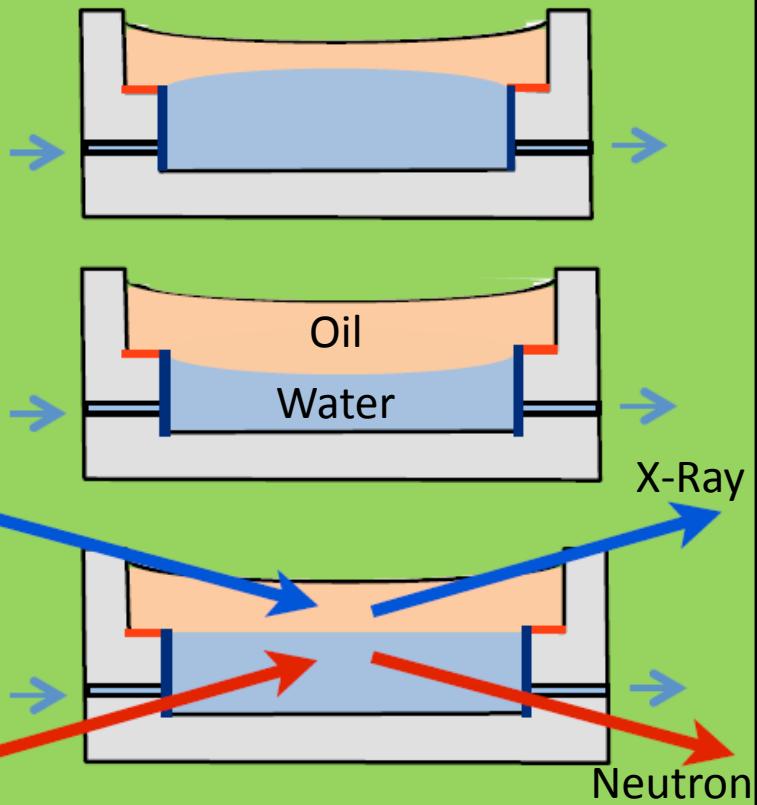
- Test measurement with neutron reflectometry at Figaro



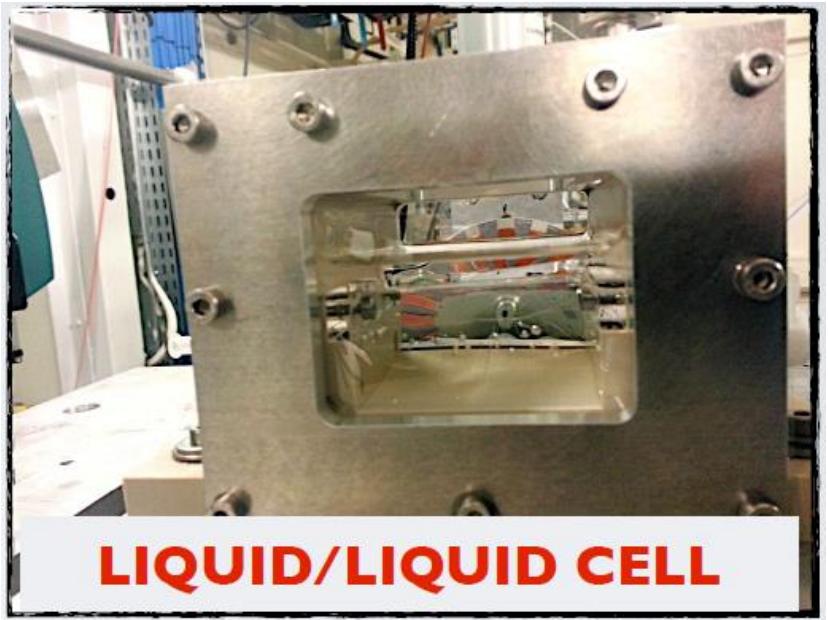
- Same substrate, but with hYeast
- First test with a peptide from cooperation partner (Jean-Philippe Michel, Paris)
- Significant change in the reflectivity curve, especially for D2O

- Lipid extraction from yeast: Find parameters for similar composition
- First sucessfull substrate preparations
- Find optimal solvent and spincoating parameters for the natural lipds
- Precharacterization with (Liquid)AFM and Fluorescence-Microscopy
- Experiment in June at DELTA (x-ray reflectivity)
- Experiment in July at FIGARO (neutron reflectivity)

Flat Interface and controlled sizes

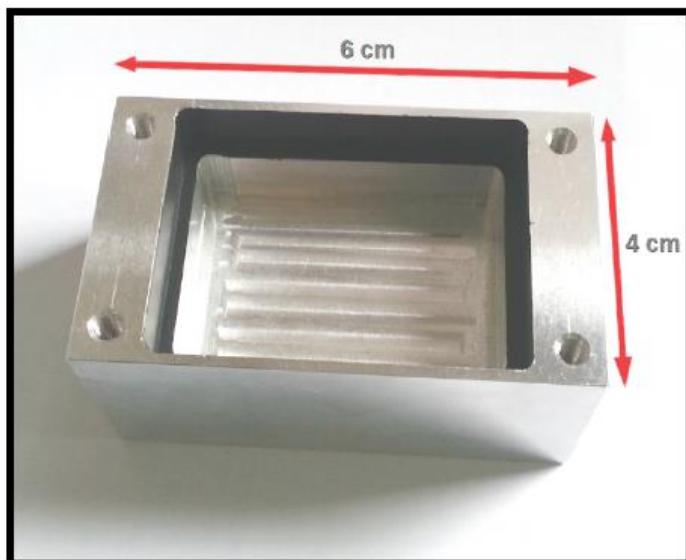


Ernesto Scoppola

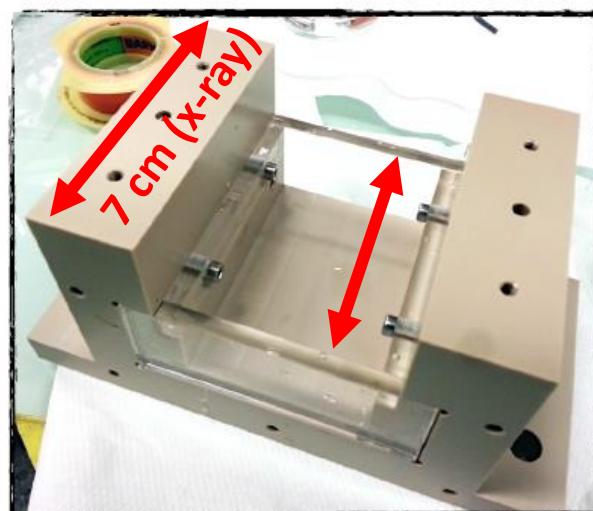
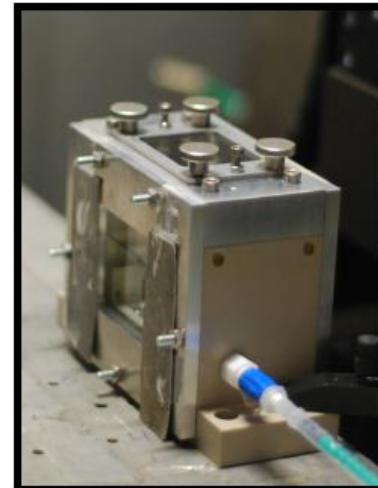


- Low Curvature with controlled sizes
- Lower Attenuation

Prototype Cell

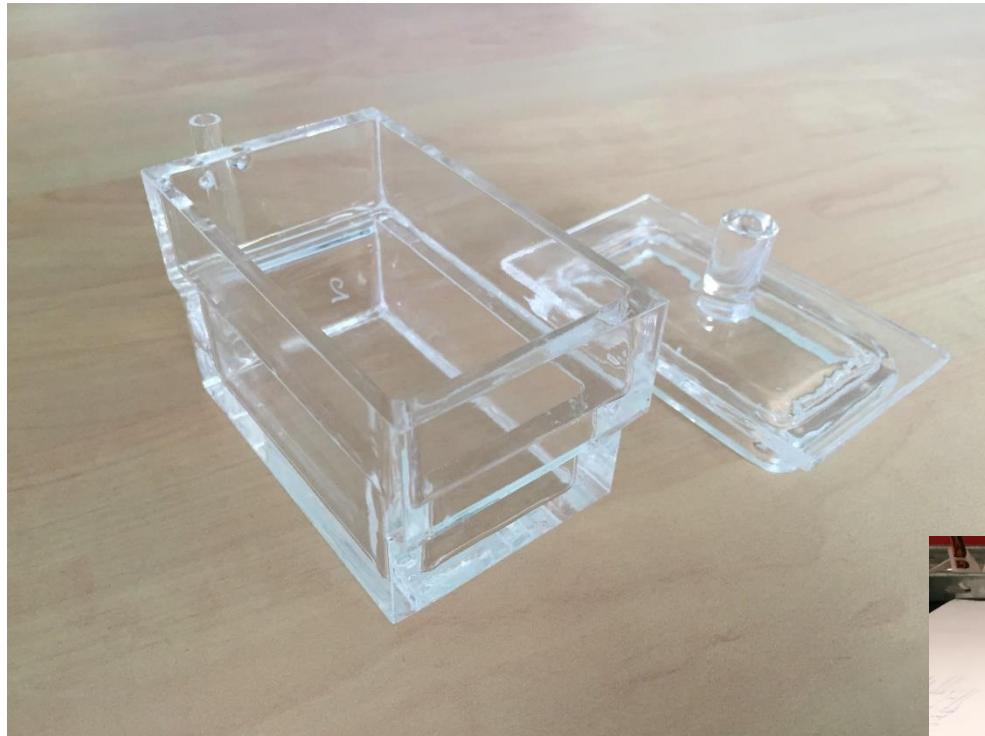


PEEK/Quartz Cell



5 cm /
3.5 cm
(neutron)

Liquid/Liquid interface



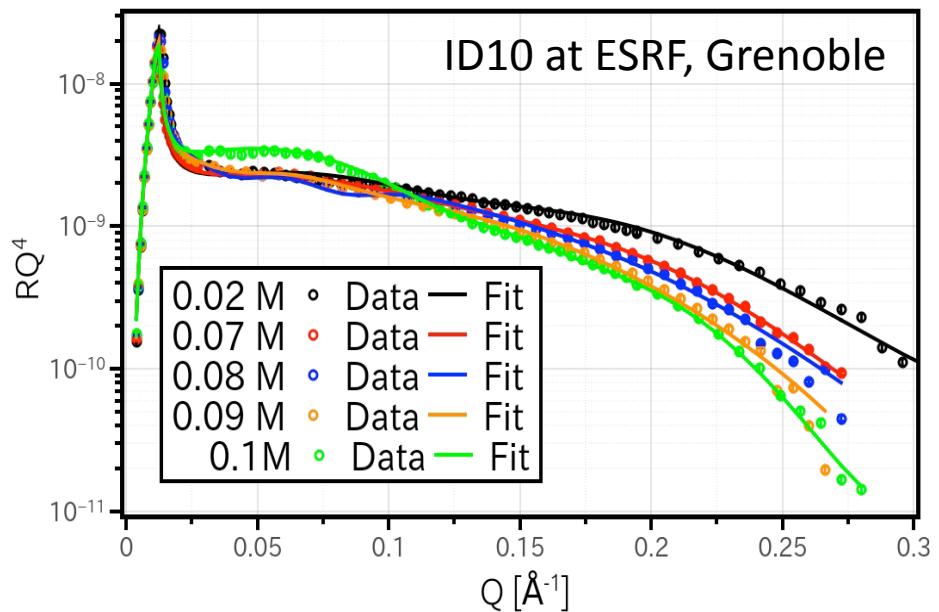
Combination of two methods:

- Neutrons: Nuclei composition (**Extractant**)
- X-Ray: Electron Density (**Ions**)

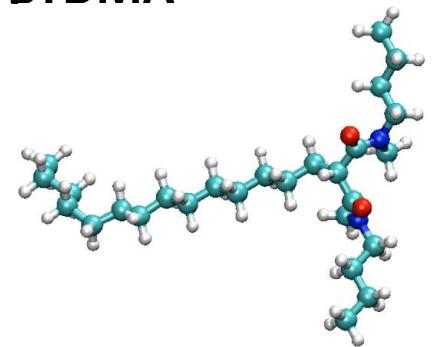
Current cells

- Quartz (One piece)
- Different sizes





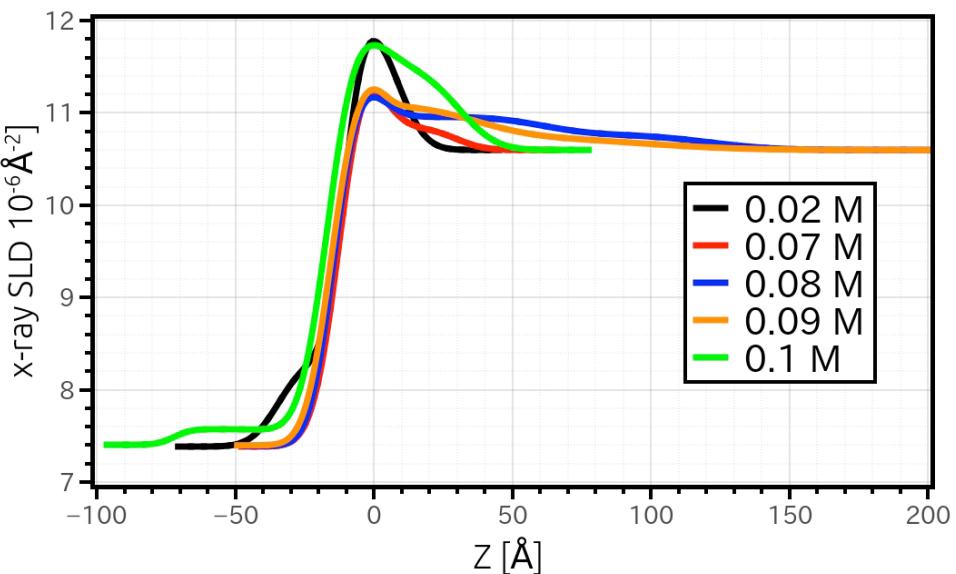
- Water with **Lithium Nitrate (2 M)** and **Neodymium Nitrate (0.25 M)**
- Dodecane
- Extractant: **DMDBTDMA**



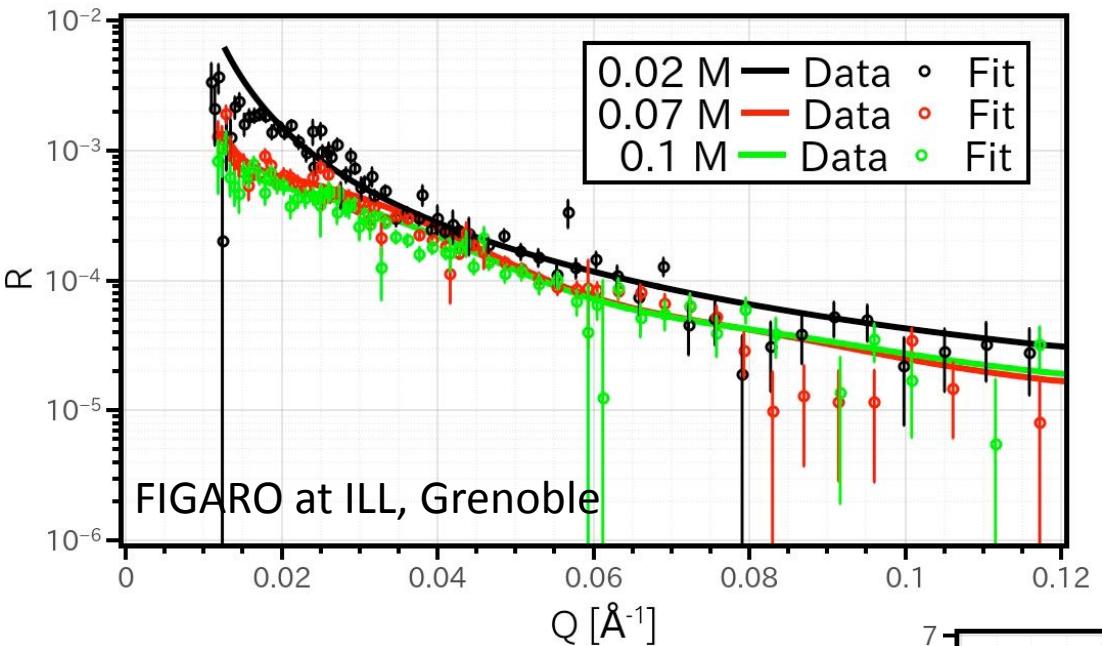
Increasing extractant concentration
 \rightarrow Change in slope

At the CAC
 \rightarrow higher intensity in the
 low Q region

Higher SLD at the interface
 \rightarrow adsorption of ions



Neutron reflectivity



Lower SLD at the interface
 → Adsorption of Diamide.

