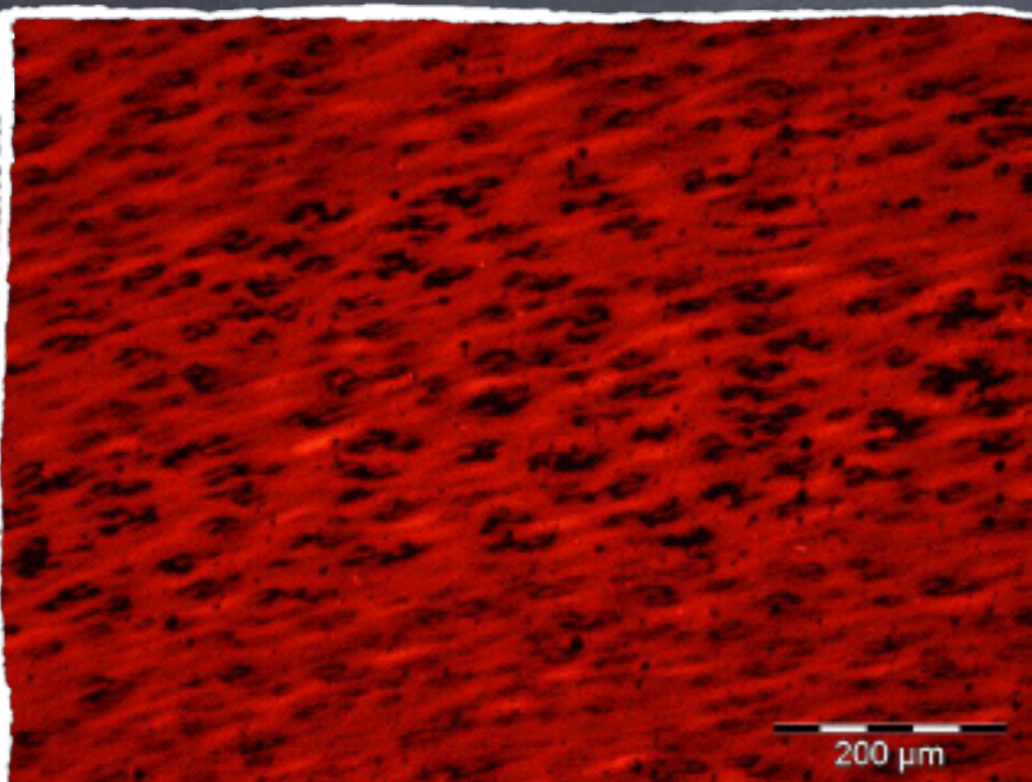
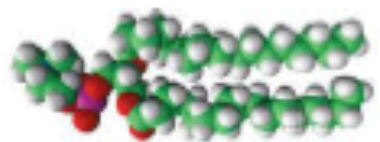


# RESULTS

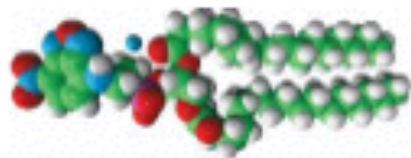


Supported Lipid Bilayer at RT

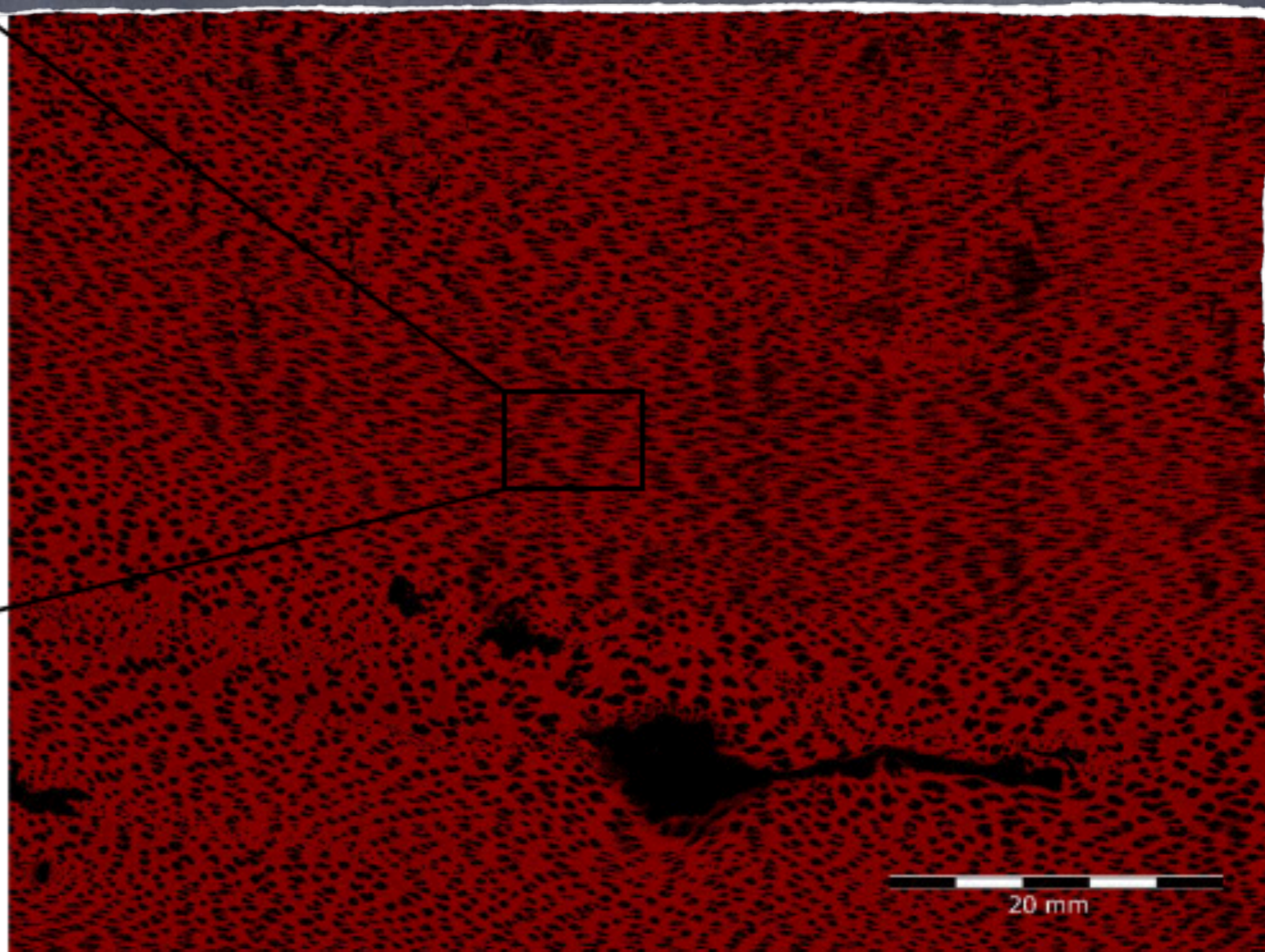


DSPC (C18)

+



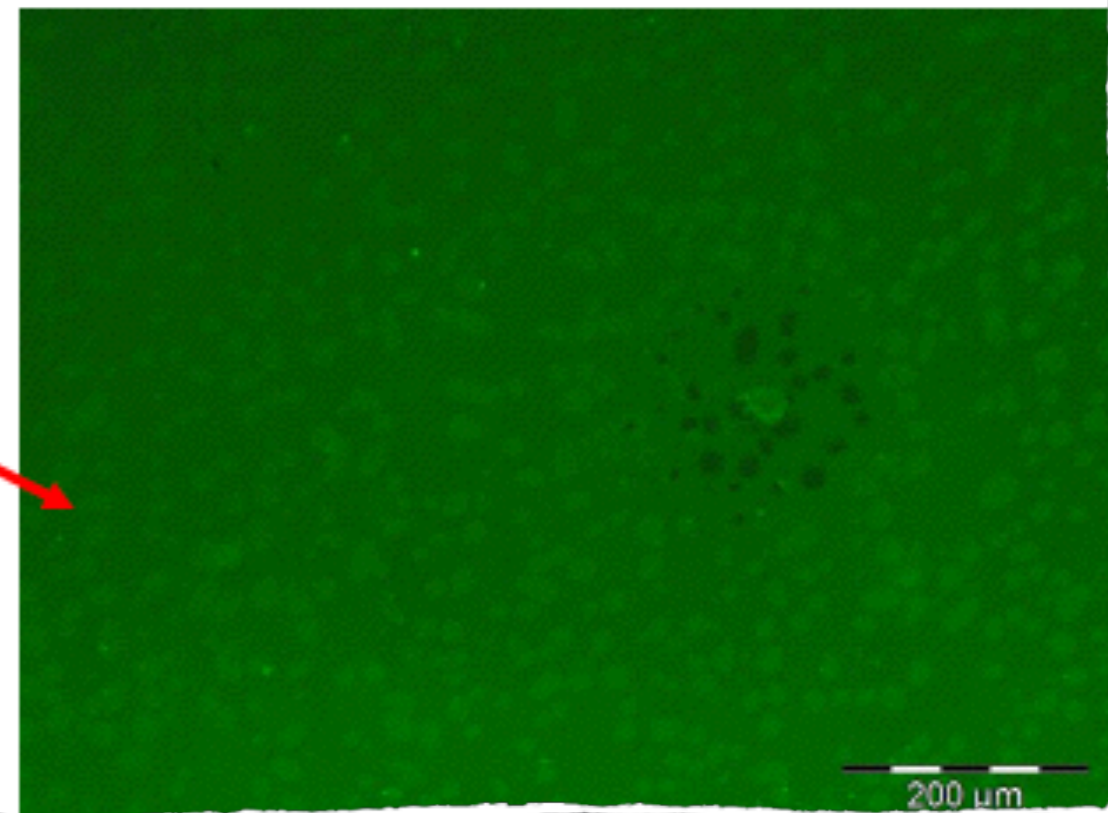
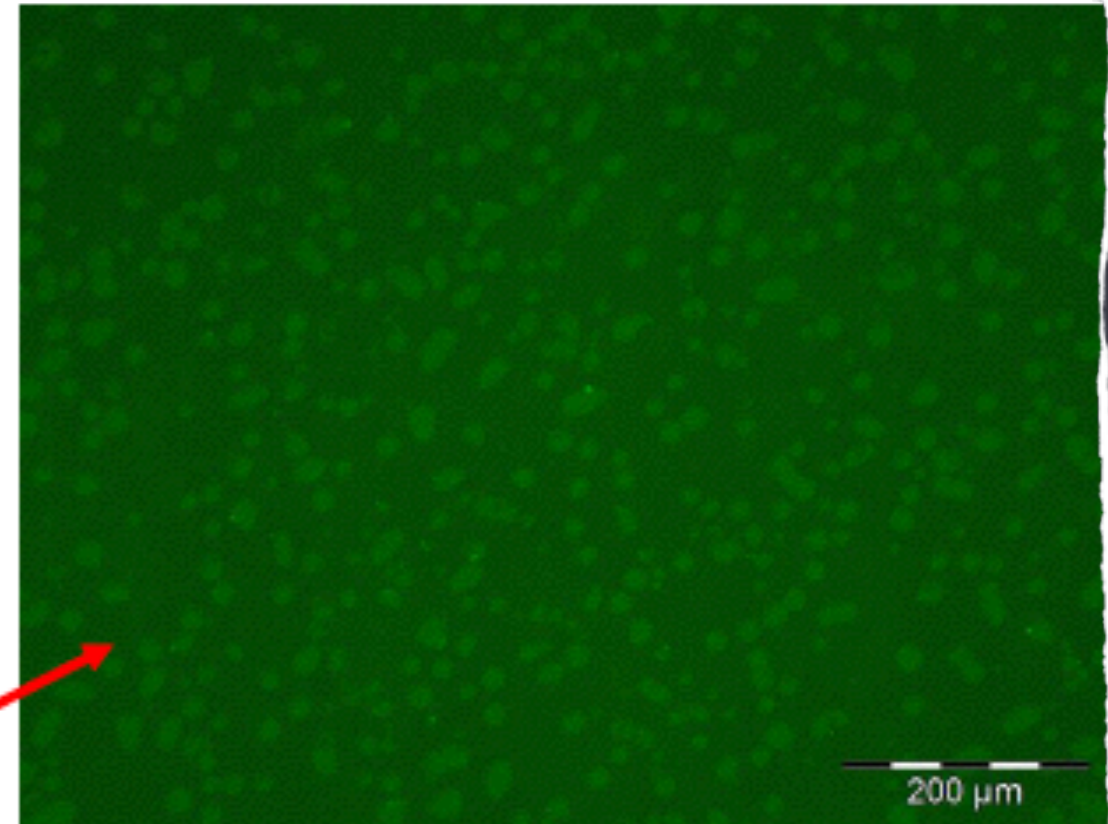
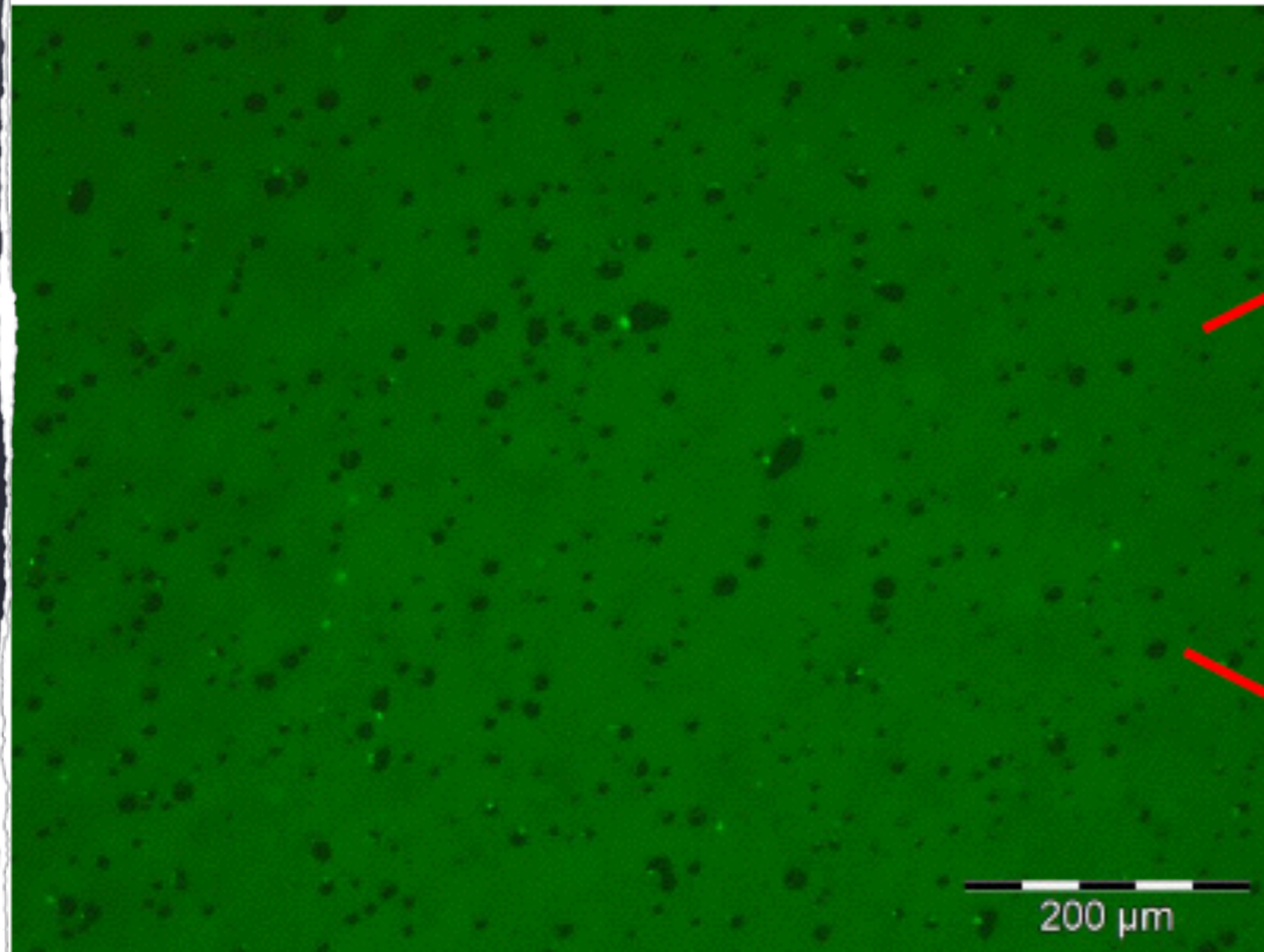
1%  
DSPE (C18)  
NBD at HG



# RESULTS

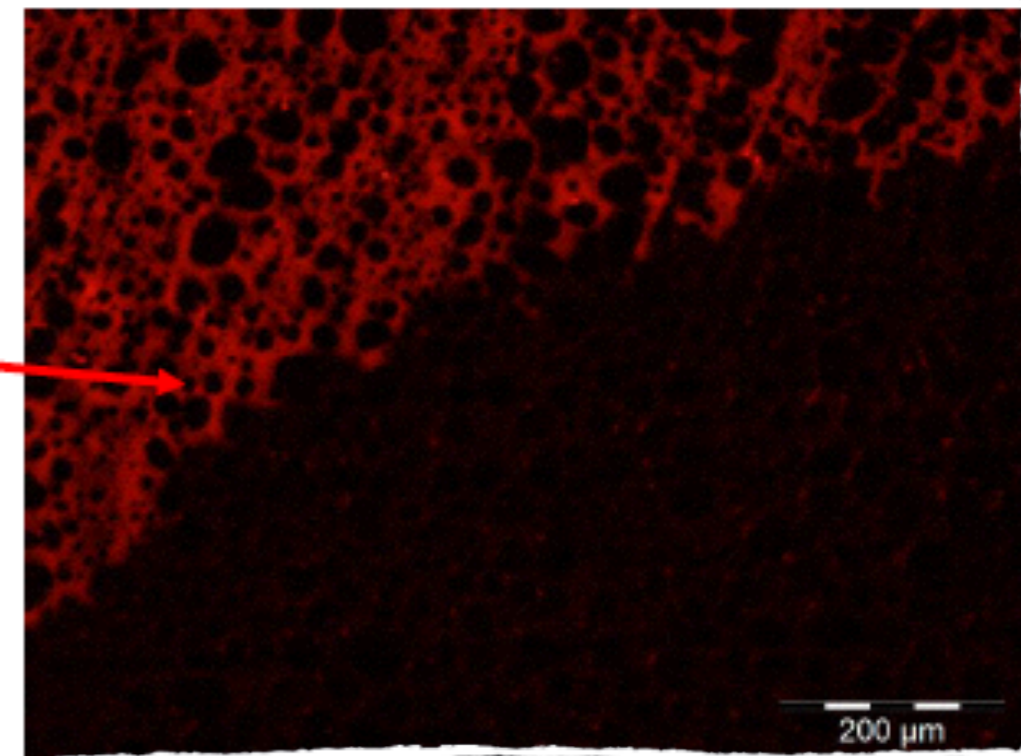
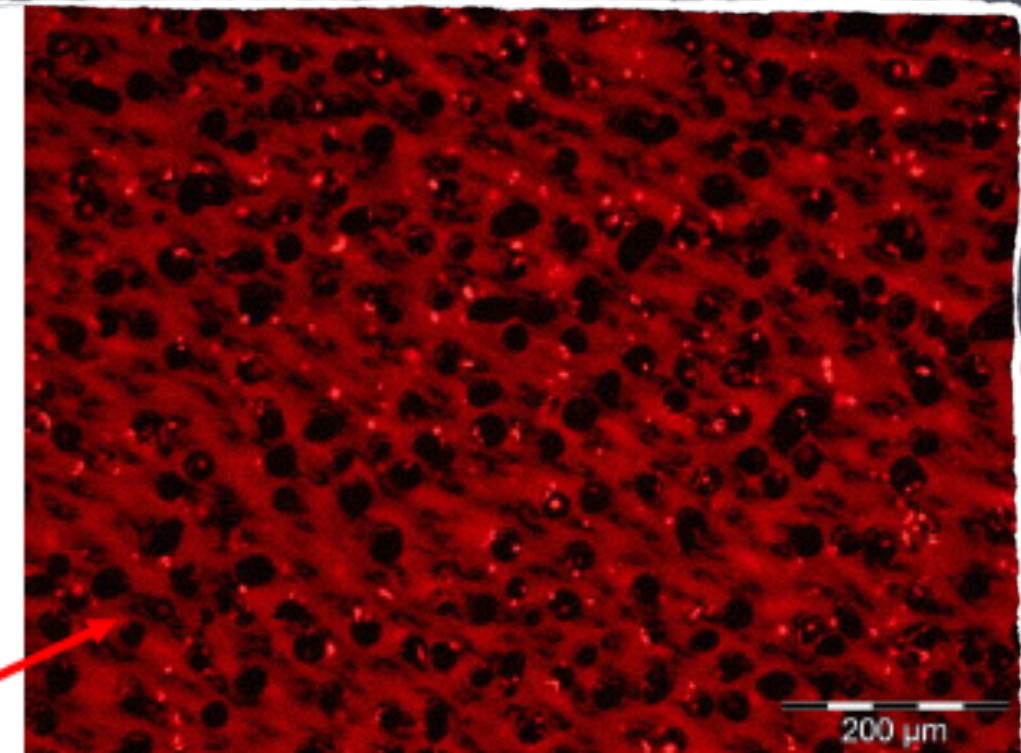
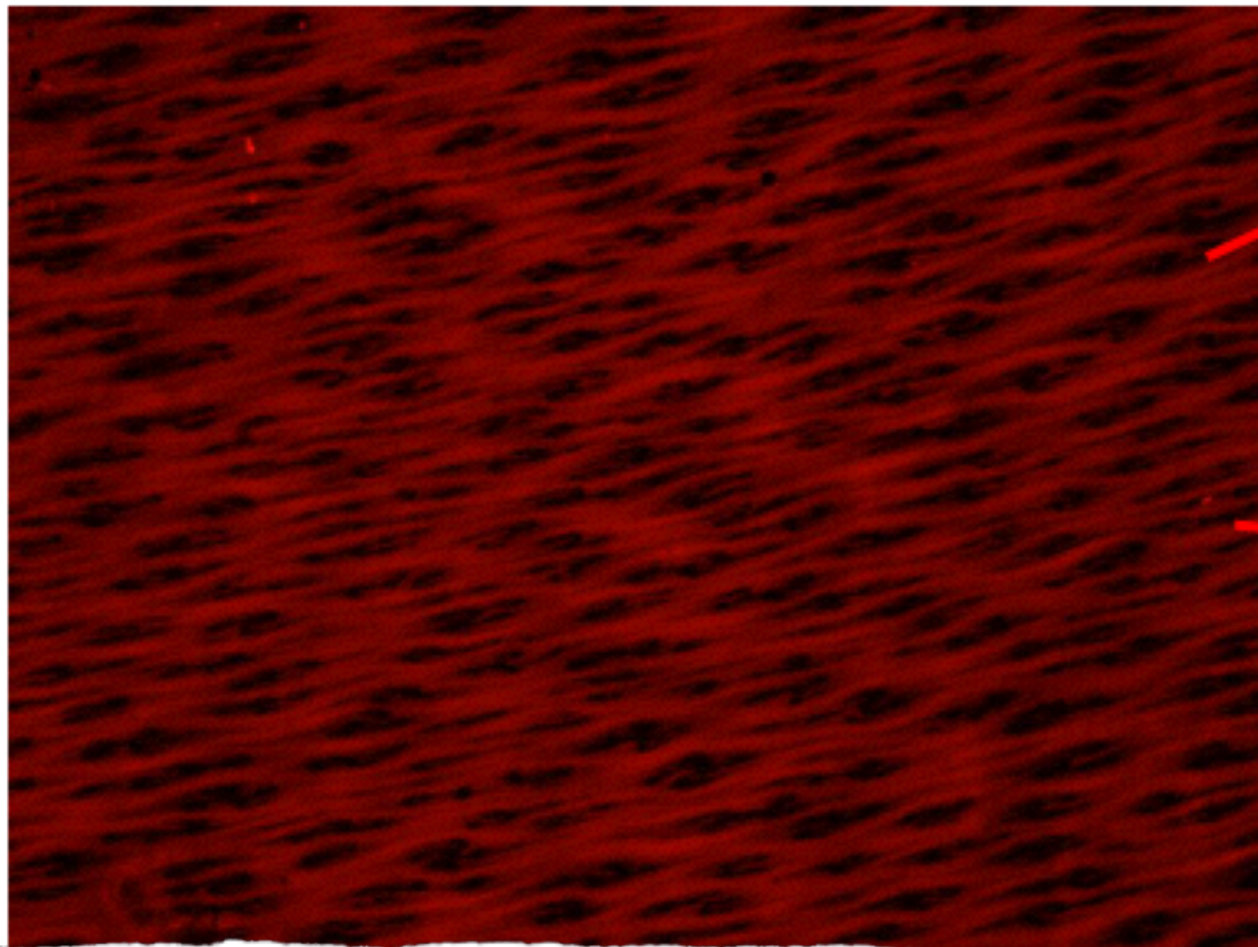
Next Step: Observe the sample in fluid phase

- First Measurement at RT
- Heat Sample for 2h at 57deg
- Go back to RT



# RESULTS

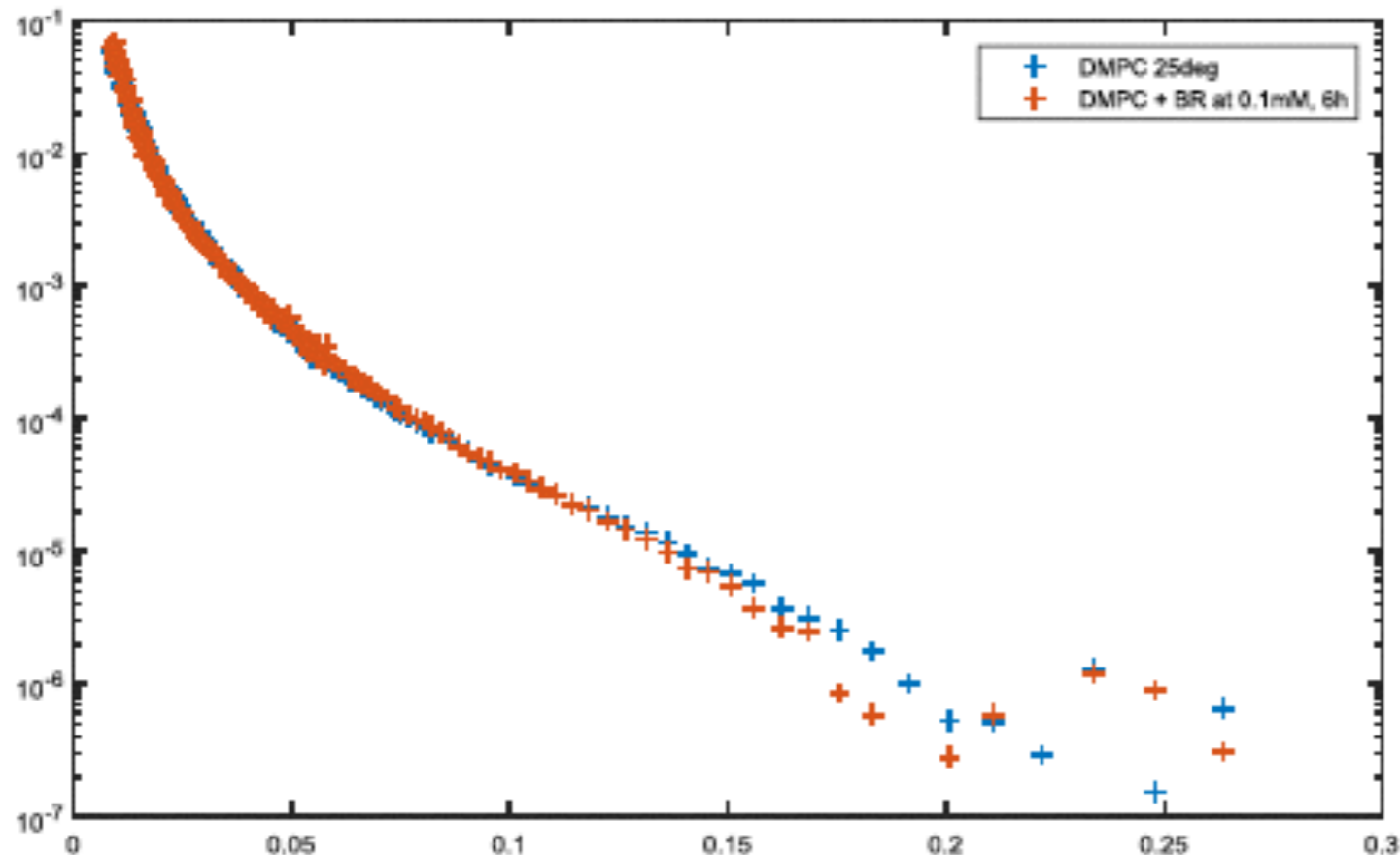
- Next Step: Saturate the membrane with detergent
  - First Measurement at RT
  - Insert detergent via mf-pump
  - Heat Sample for 2h at 57deg
  - Rinse detergent
  - Go back to RT



# RESULTS

## Recently Neutron Reflectivity Experiment at D17

- Much more stable bilayers on silicon
- Saturation with detergent and heating much easier
- Follows the complete Protein-Insertion procedure
- Using deuterated DMPC Lipids and undeuterated Proteins
  - -> Insertion should be visible



- DMPC Sample, incubated for 6h with BR
- Indication of protein insertion
  - Changes in the high q
- Data treatment ongoing

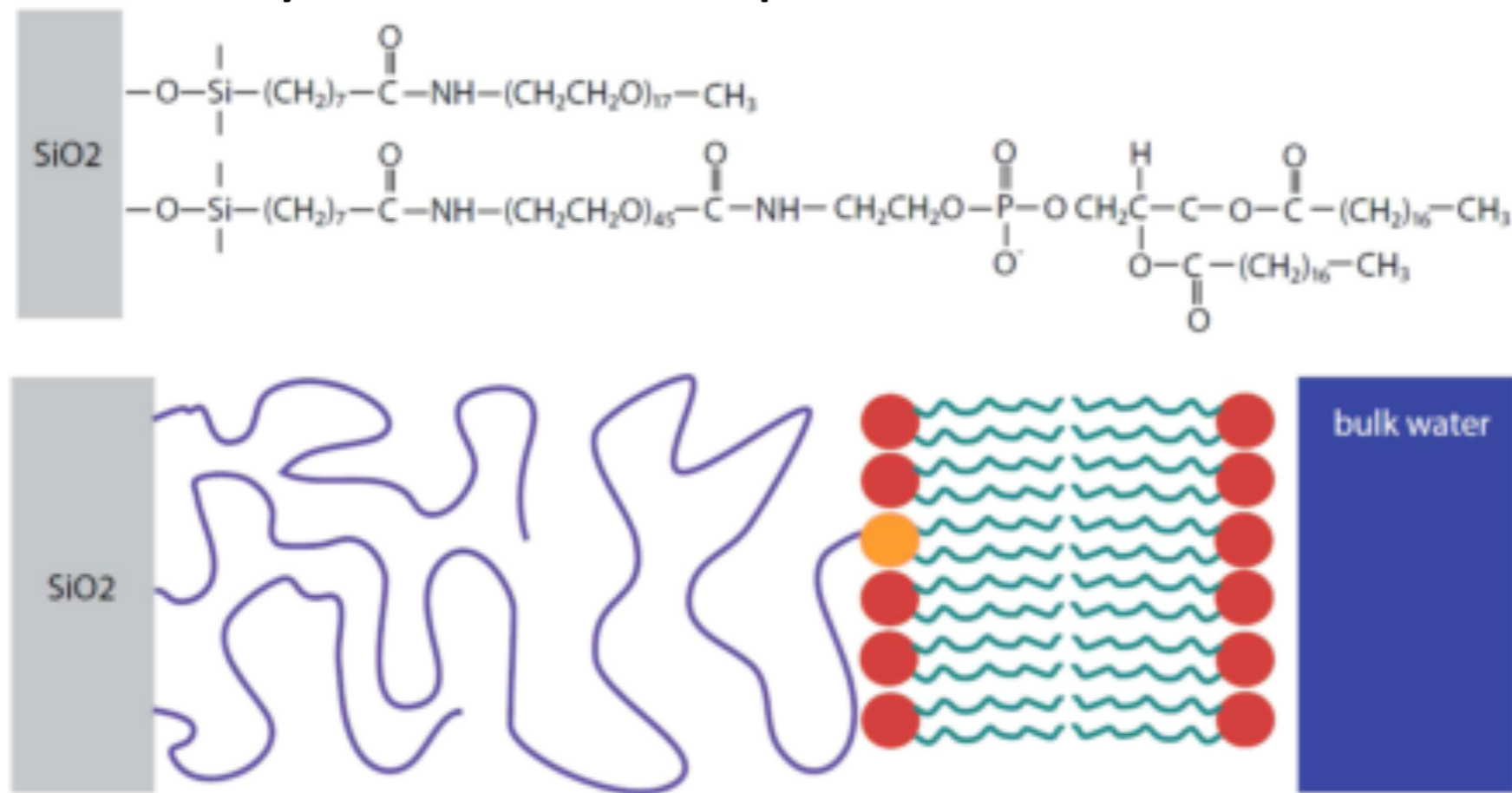
# CONCLUSION OF WORK STILL IN PROGRESS...

- Sample System to observe lipid bilayers with fluorescence microscopy is developed
  - Available at the PSCM!
- Bilayer coverage and stability seems to be inferior compared to silicon
  - Accomplishing successful protein insertion quite difficult
  - Optimization: Another substrate (polished quartz)
- Neutron reflectivity experiment
  - Indications of successful protein insertions could be seen

## Collaboration with AG Nickel from Munich: Polymer-tethered spin-coated membranes



Irena Kiesel



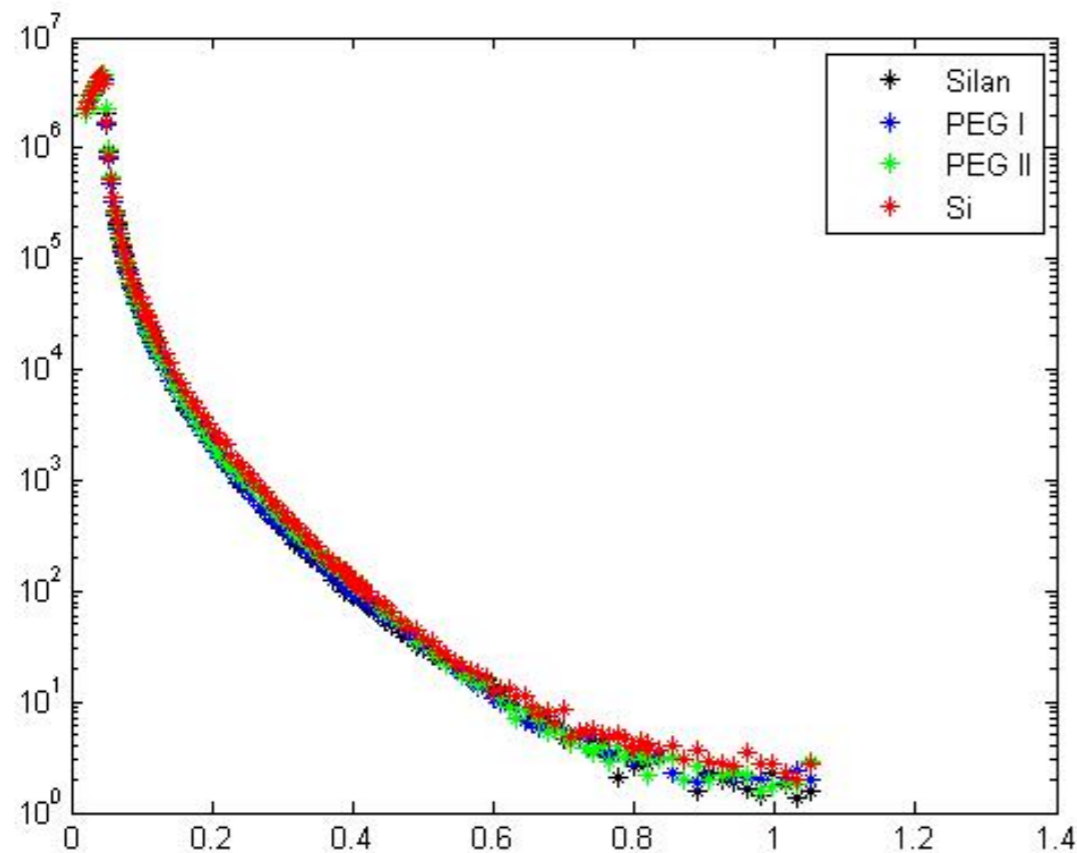
Hertich et al.,  
Langmuir 2014, 30, 9442-9444

- Short and long polymer brushes (PEG) as tether for membranes, chemically grafted on silicon
- Deuterated and natural lipids extracted from yeast (de Ghellinck, current project PhD Robin Delhom) to mimic natural membranes

Advantage:
 

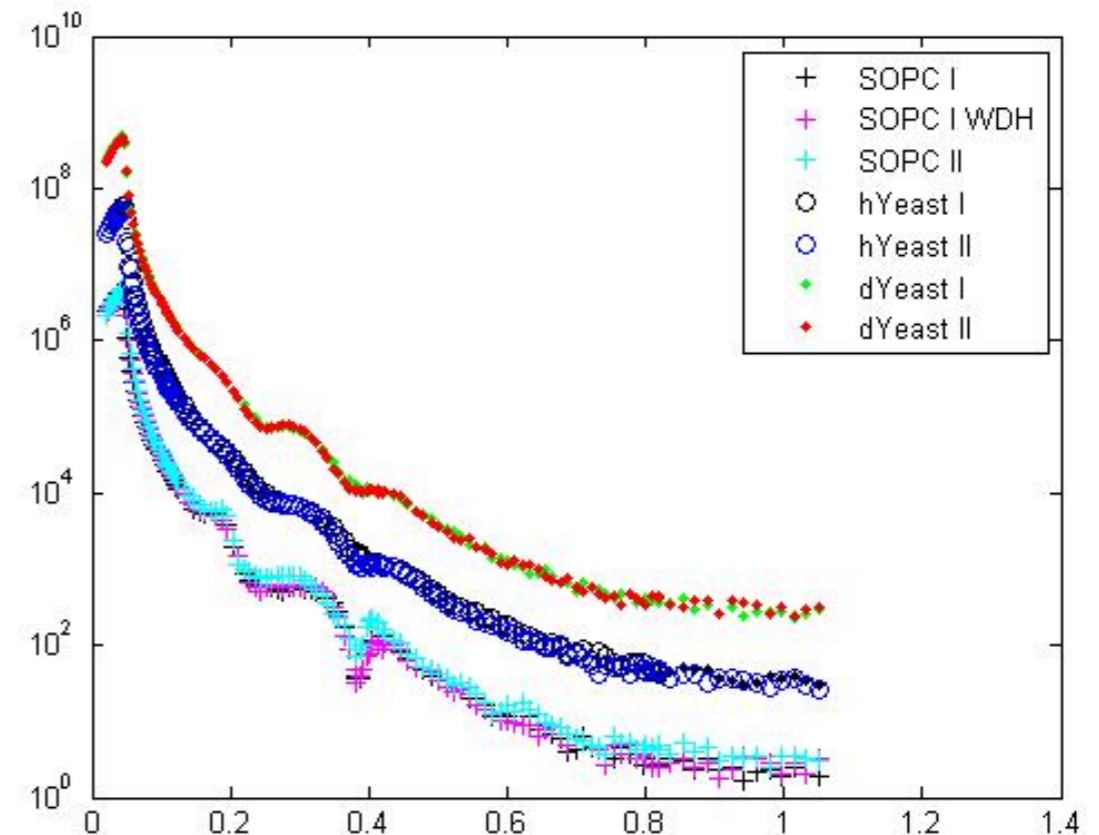
- Fast easy sample preparation with spin-coating or solvent exchange
- Reusable substrates

courtesy I. Kiesel



## Substrates in water

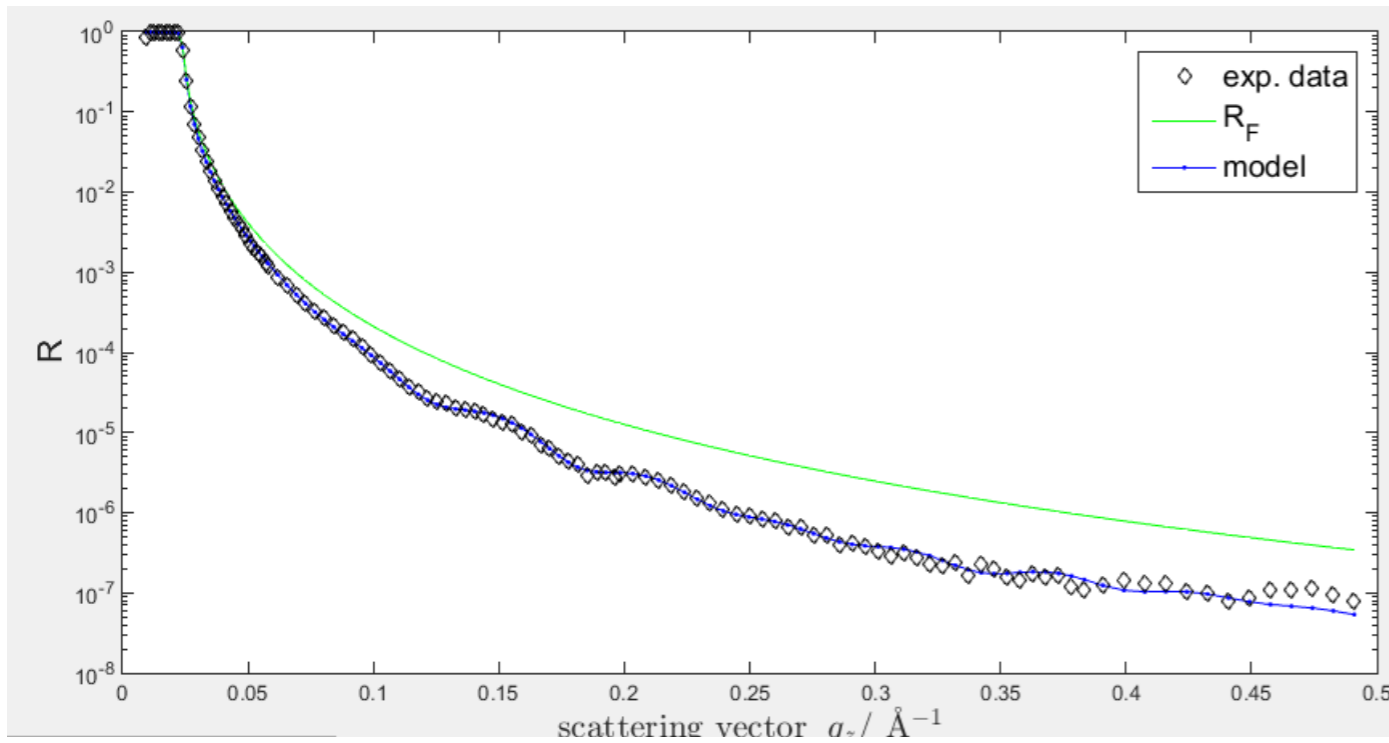
- Clean silicon (red)
- With Octenyltrichlorosilan (black)
  - Nearly not visible in contrast to OTS (hydrophobic gap?)
- Two substrates with PEG brushes (I blue & II green)
  - PEG brushes not visible in water
  - same electron density



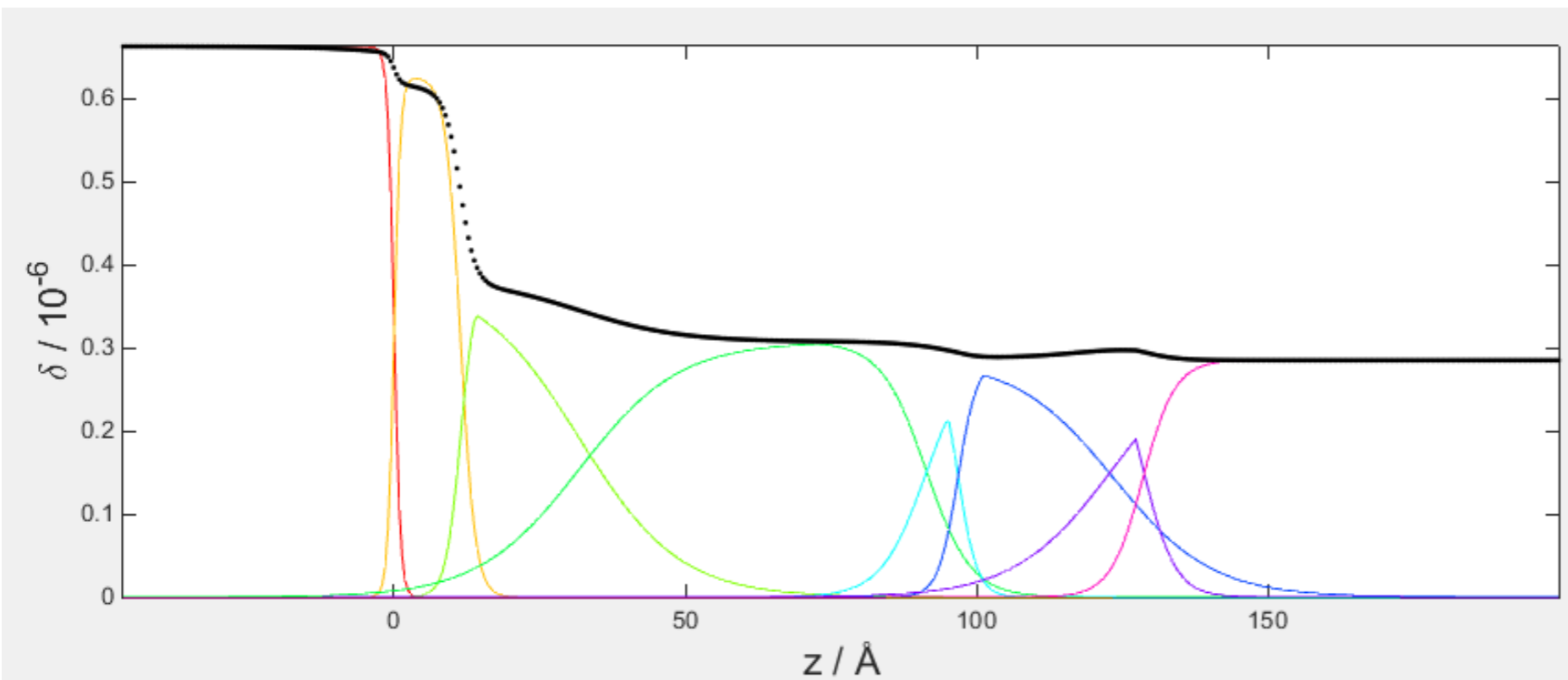
## Spin-coated lipids on PEG brushes substrate I and II

- cleaned with chloroform and methanol in between)
- Reproducible with the same lipid
- With SOPC: stronger features → more ordered (?) than natural mixture of lipids
- H-Yeast and D-Yeast similar

courtesy I. Kiesel



- H-Yeast on PEG brushes
- PEG brushes denser at the interface
- Low contrast between PEG and lipids
- Interface still a bit too rough to distinguish good between head- and tail-groups

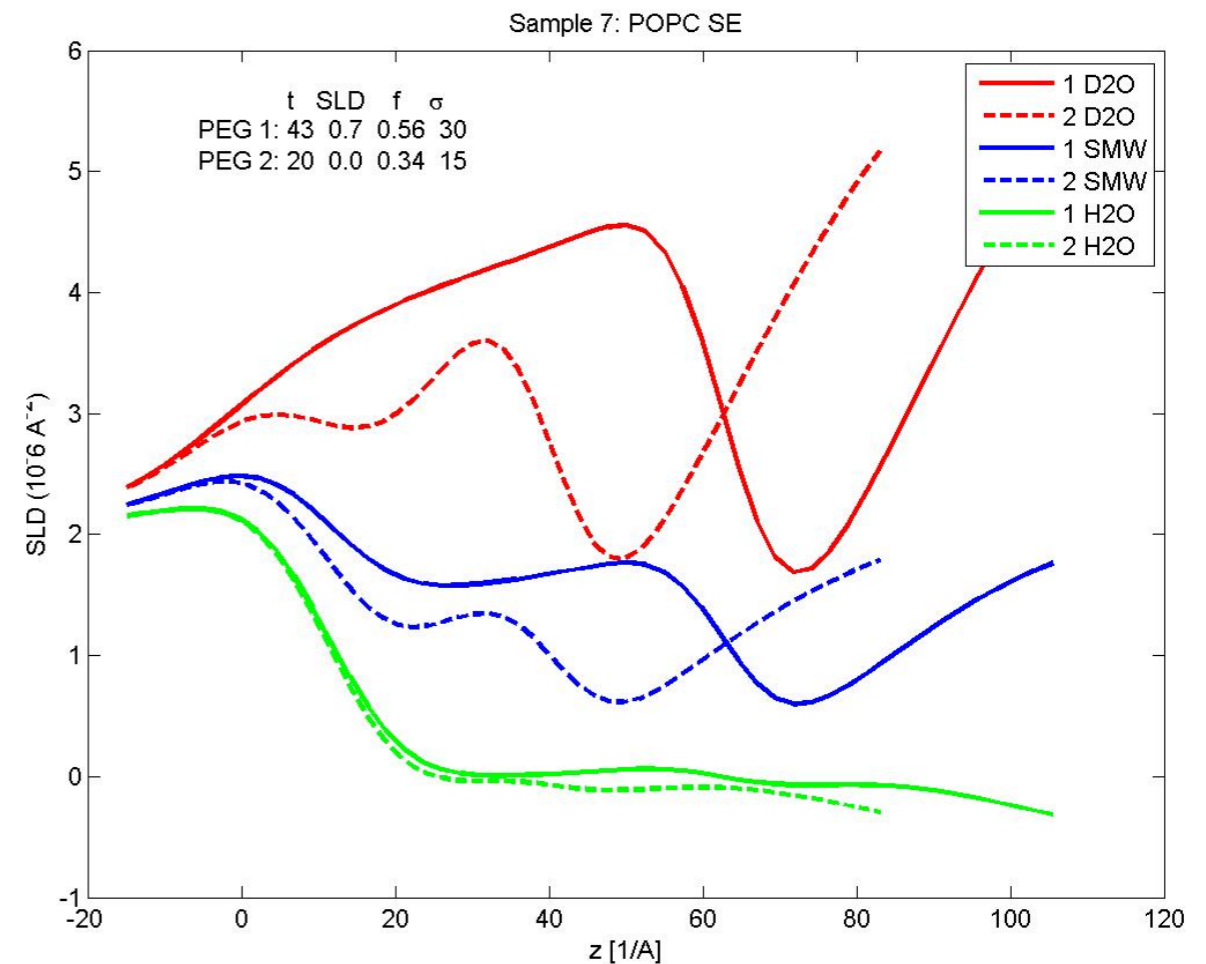
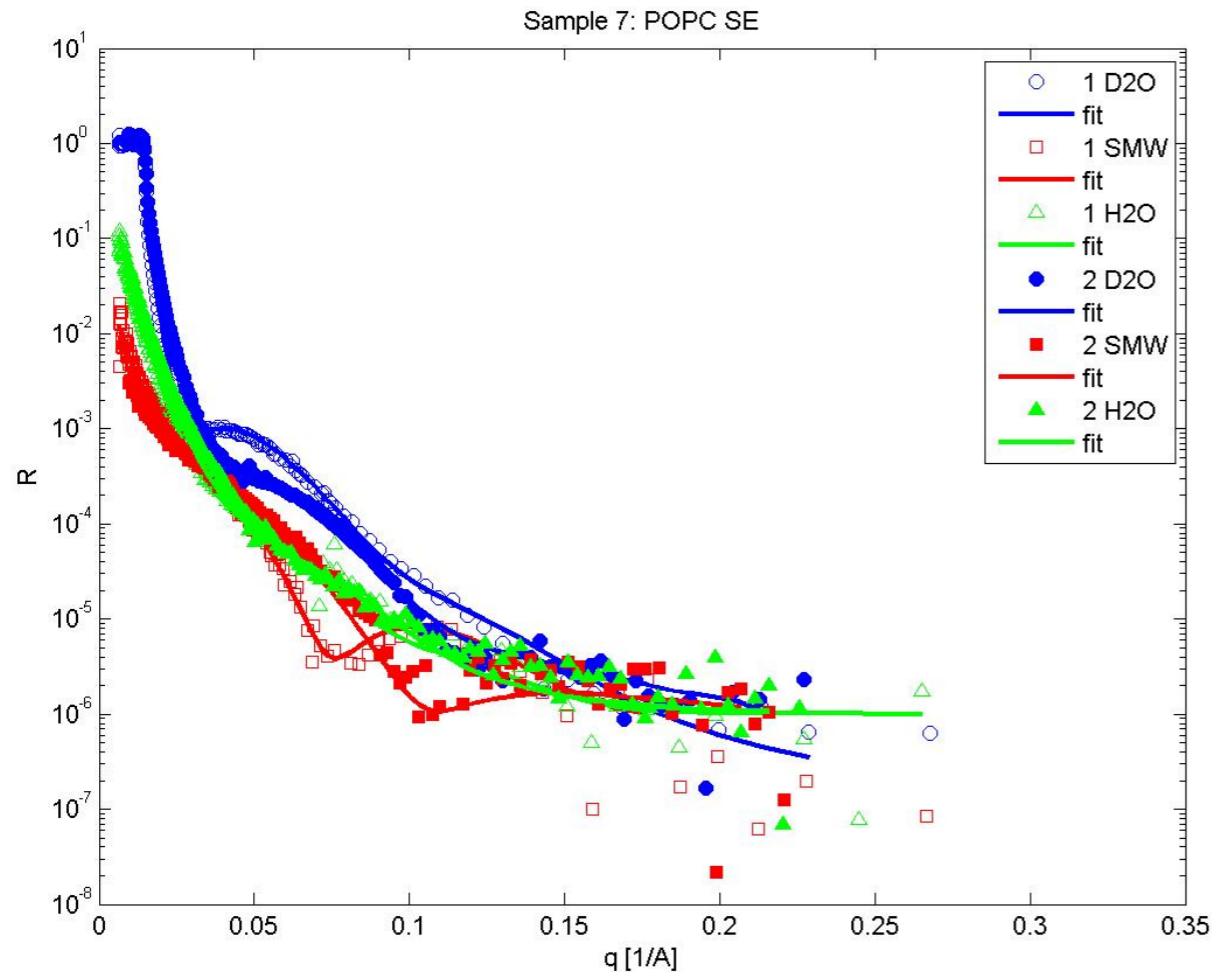


Si      SiO<sub>2</sub>      PEG      Lipid-Bilayer      Water

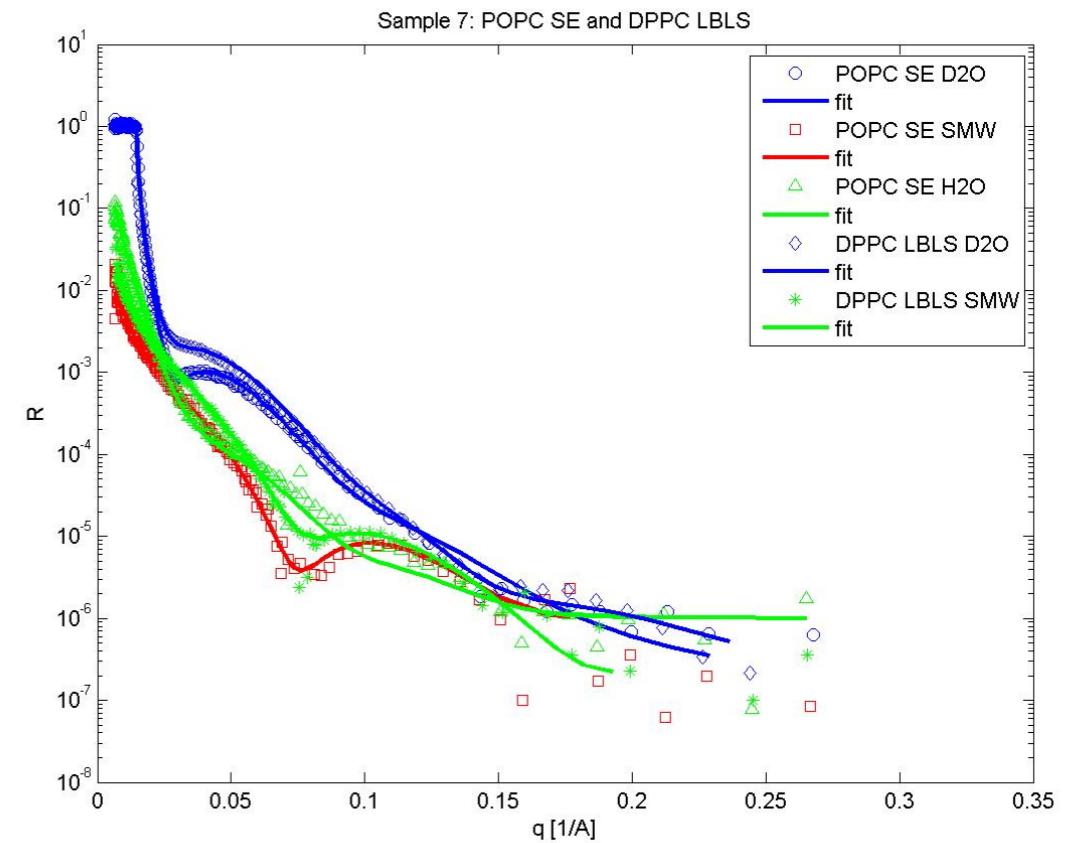
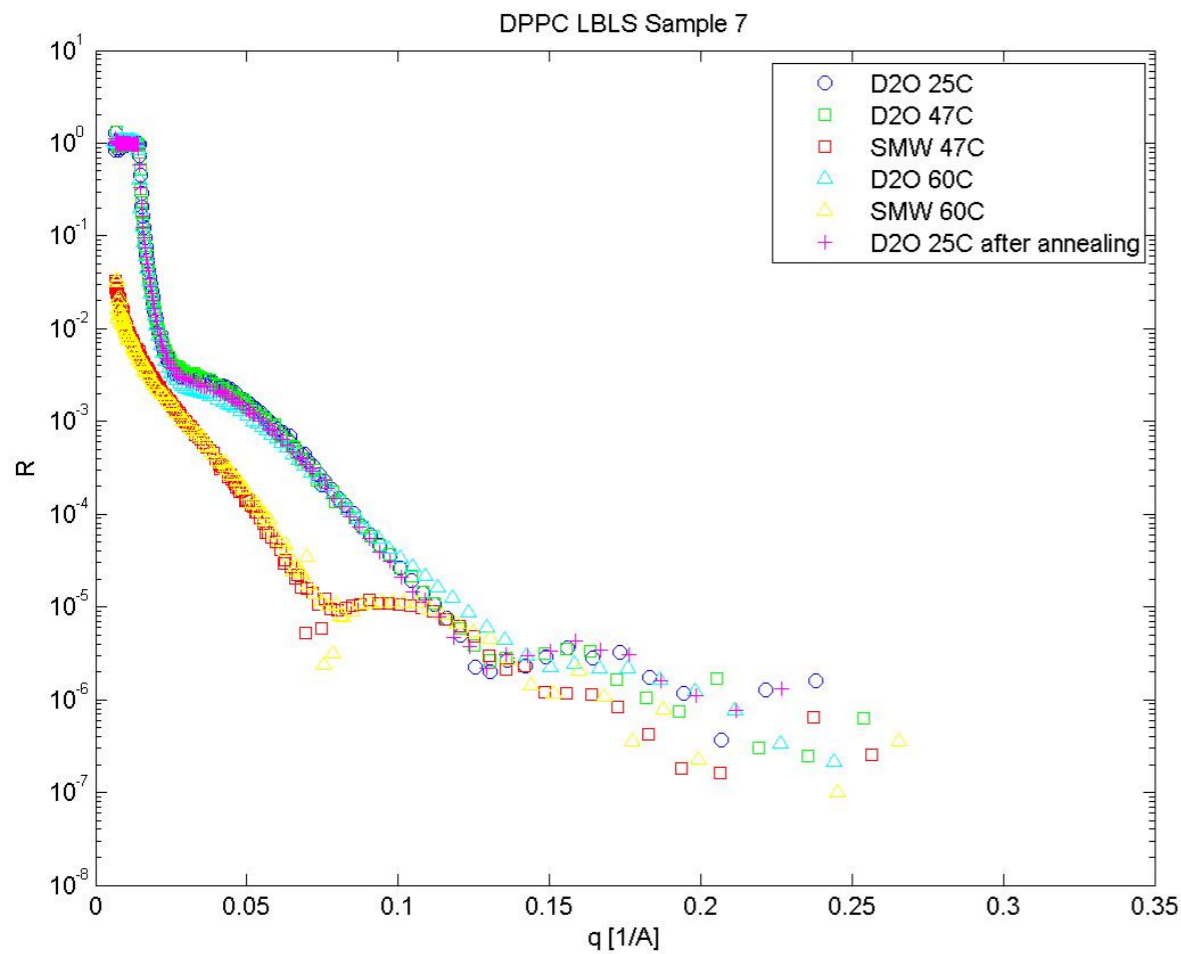
courtesy I. Kiesel



Test of the reproducibility with different deposition methods: solvent exchange, spin coating, vesicle fusion, LB-LS

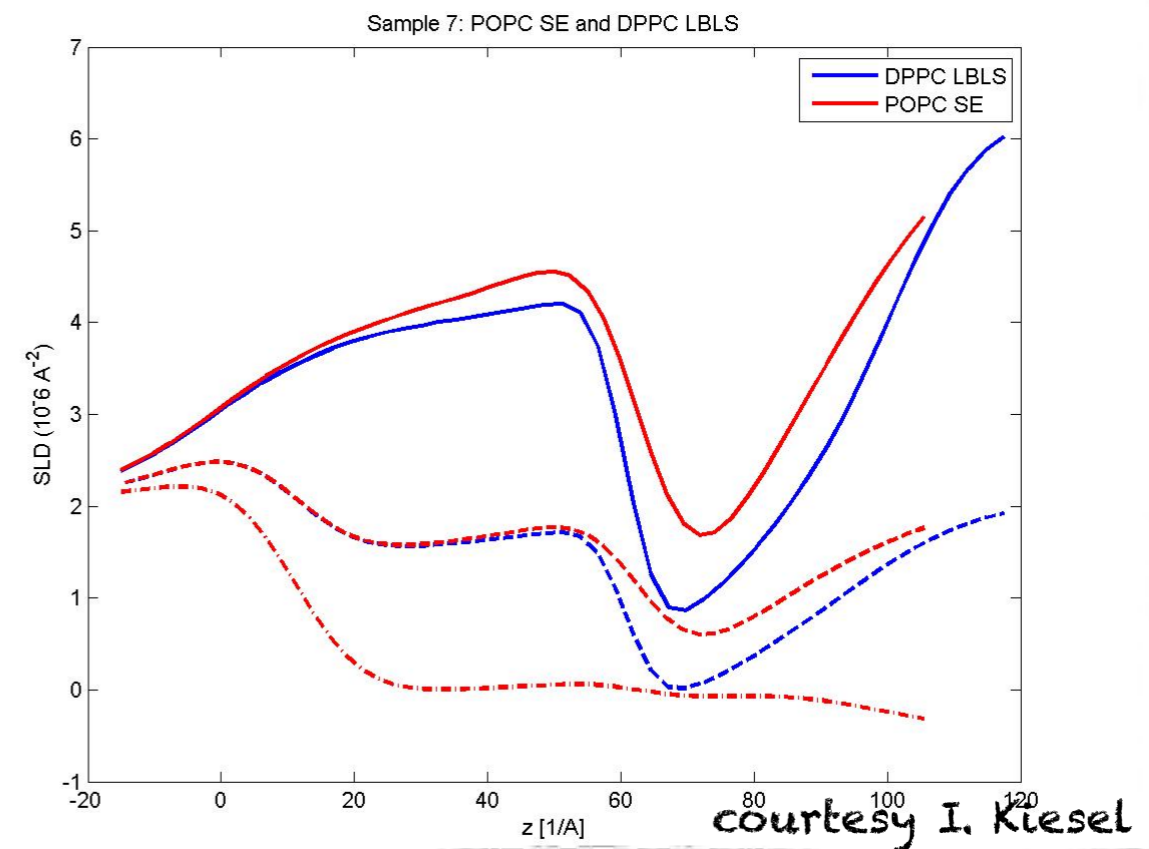


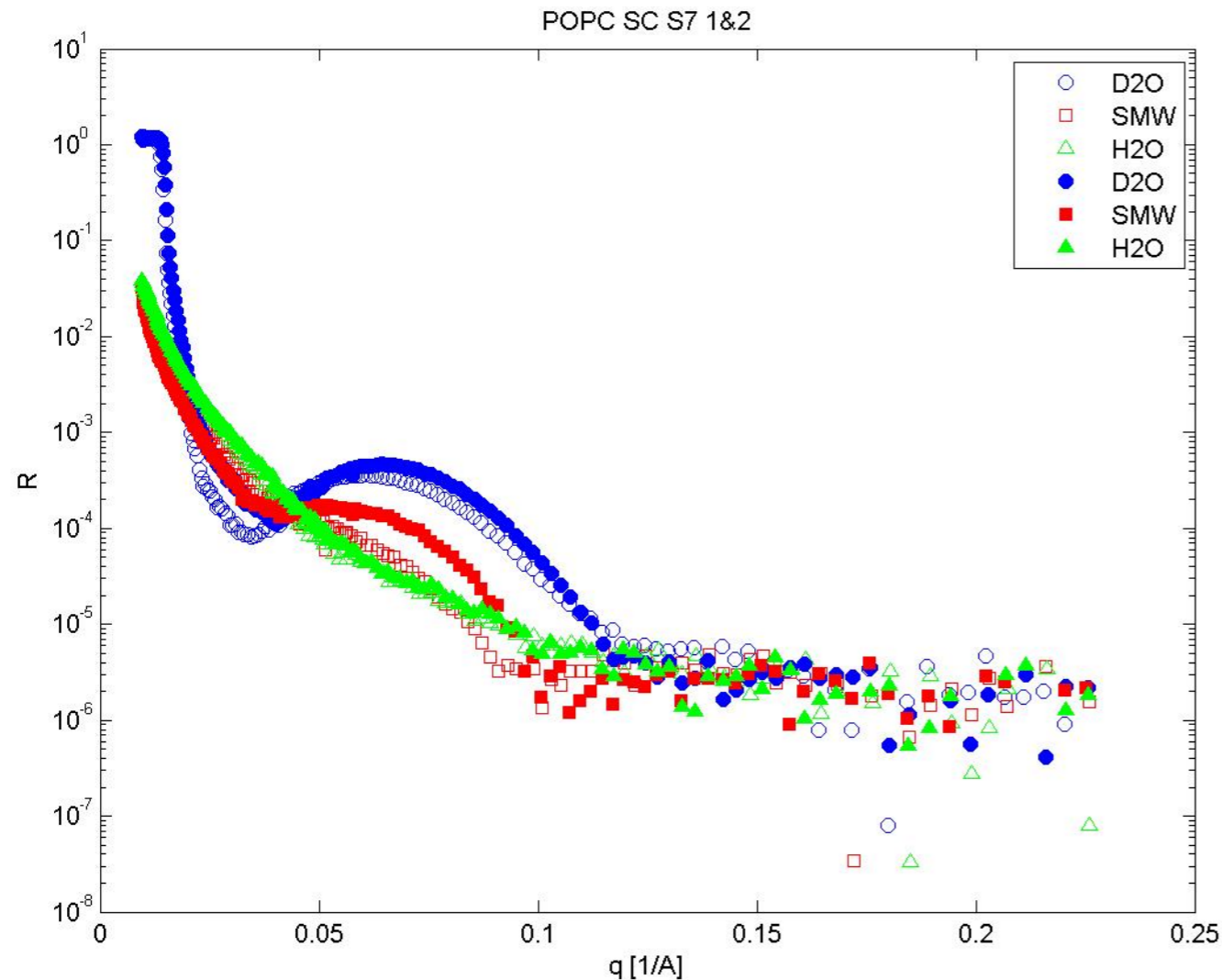
- Same substrates at different experimental times
- Solvent exchange (after cleaning with methanol of a spin-coated lipid layer, measured already a couple of hours in water)
- Reflectivities not similar!
- But: The lipid bilayer is the same, only the PEG layer changes
  - Does it need a long time to swell in water?
  - Current tests with ellipsometry and long measurement at FIGARO at the end of this week



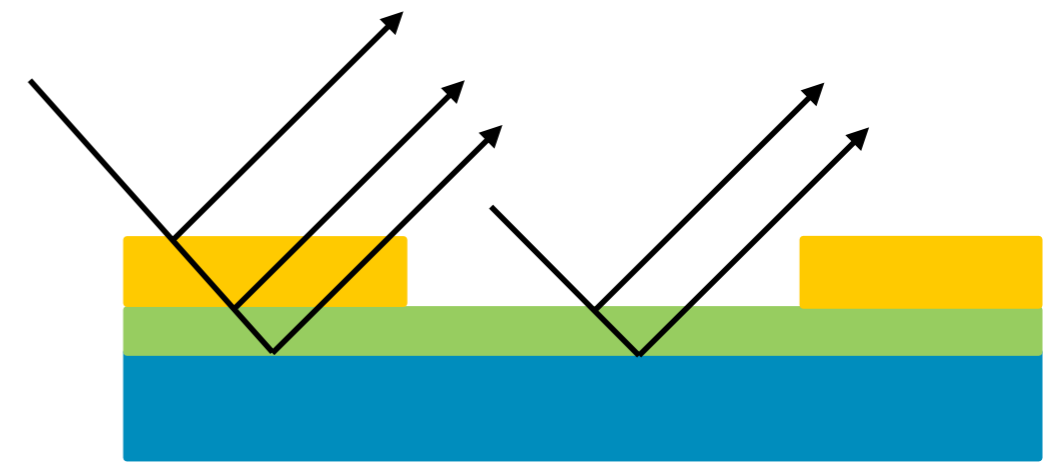
DPPC with LB-LS on same PEG substrate

- Heating to different temperatures for annealing
  - Nearly no change in reflectivity
- Comparison LB-LS/solvent exchange
  - Comparable roughness and thickness, slightly different density (different lipids, coverage?)

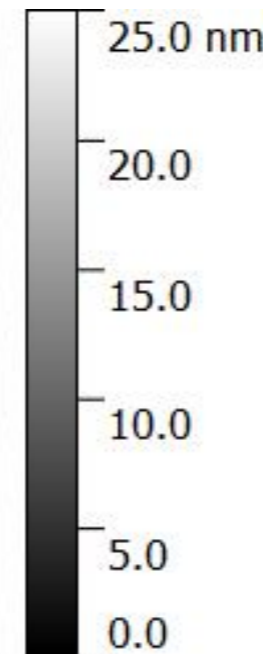
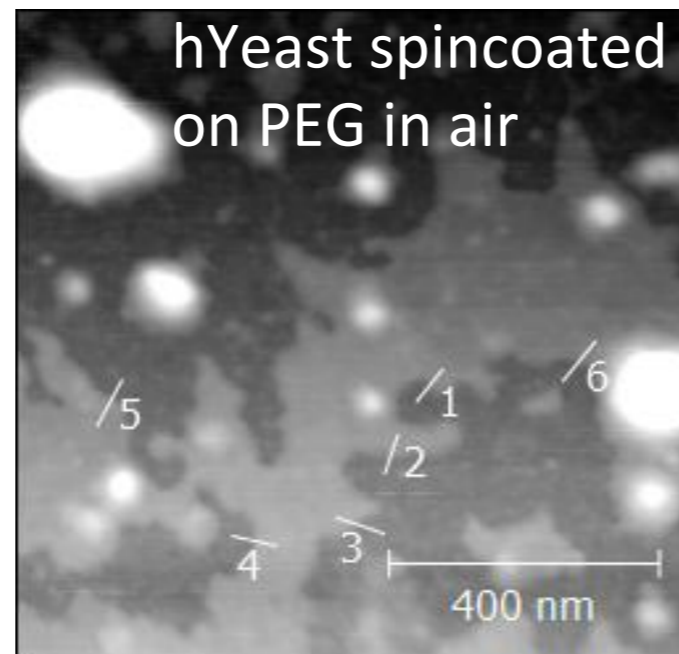
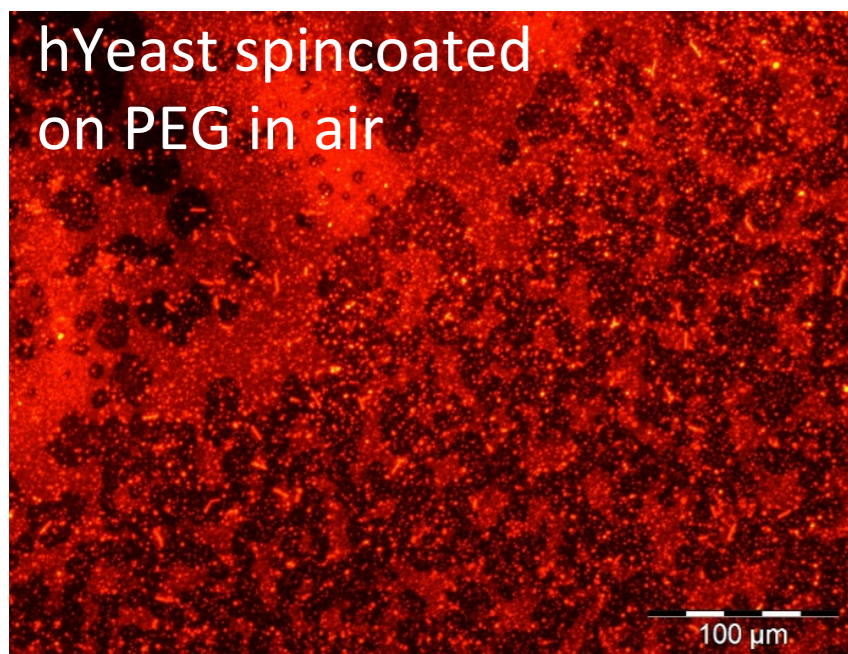




Repeated spin-coated POPC on PEG  
 Broad minima: Difficult to fit → Patches?



- Check with fluorescence microscopy and AFM
- Optimization of spincoating parameters



Spin Coater Delta6 SUSS MicroTec

courtesy I. Kiesel

- Possibility to create stable lipid bilayer with different techniques on PEG brushes (vesicle, spin-coating, solvent exchange, LB-LS)
- Reproducibility still a problem, maybe due to swelling time after hydration
  - No problem with XRR
    - Smaller samples
    - lower sensitivity?
    - luckily measured at the same time after preparation and hydration?
- Optimizing of spin-coating parameters
- Solvent exchange seems even better than spin-coating
  - cleaning and preparing directly at the instrument possible
  - positively tested with natural lipids as well

